Syntheses of Glucose-Containing Lipid A Analogues and Their LPS-Antagonistic Activities

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Three anomeric pairs of lipid A-type disaccharides containing a glucose on their reducing end were synthesized, and their LPS-antagonistic activities were measured. The inhibitory activities (IC₅₀) on the LPS-induced TNF α production of these six compounds (16 α , 16 β , 28 α , 28 β , 40 α , and 40 β) toward human whole blood cells were 0.35, 0.42, 2.37, 1.16, 2.89, and 7.70 nM, respectively.

The study of endotoxin, which was named by R. Pfeiffer in 1892,1 has developed extensively2 since Shiba and Kusumoto's³ total synthesis of lipid A, a toxic component of endotoxin (lipopolysaccharide, LPS), existing in the outer surface membrane of Gram-negative bacteria. In an earlier investigation to search for drugs, lipid A-related compounds were investigated as anticancer drugs² by stimulating the immune system.4 Conversely, a nontoxic natural lipid A-related compound (RsDPLA)⁵ was recently isolated from *Rhodobacter* sphaeroides by an Eisai group. This compound, unlike lipid A, has a unique structural feature, that is, it contains a cisor trans-alkene and a 3-oxoalkanamide in one of its long fatty acid chains, and does not show LPS-agonistic activity toward mouse macrophages.⁶ Furthermore, the Eisai group found that many RsDPLA-related compounds having an olefinic double bond in their molecules behave as LPS antagonists toward both human and murine macrophages,⁶ and E5564,⁷ a compound related to RsDPLA, has been developed as a highly potent anti-septicemia drug.

The active structures of natural RsDPLA and synthetic RsDPLA-related compounds, such as E5564, are constructed with an $\beta(1-6)$ linked glucosamine–glucosamine disaccharide moiety, and the configuration of the anomeric position of the reducing end is α without exception. Regarding this structure, we addressed two questions: (1) When one of the glucosamine parts or both glucosamine moieties are replaced with a glucose or two glucose molecules, respectively, do these compounds still retain the LPS-antagonistic activity? (2) When the configuration of the anomeric position of these unnatural glucose-containing compounds is β , do they still keep the LPS-antagonistic activity?

To examine these questions, at this time, some compounds in which the non-reducing part was the same glucosamine as that of E5564 and the other reducing end was a glucose were synthesized, and their biological activities as LPS-antagonists were measured. This paper describes the synthesis of six com-

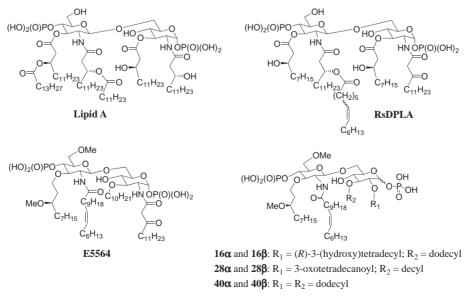


Fig. 1. Structures of Lipid A, RsDPLA, E5564, and six synthetic compounds.

Scheme 1. Reagents and conditions: a) dodecyl methanesulfonate, NaH, DMF, rt, 16 h, 75%: b) allyl alcohol containing 2% HCl, reflux, 20 min, α : β = 10:1, 80% (3α), 8% (3β); c) 2,2-dimethoxypropane, p-TsOH, DMF, rt, 16 h, 81%; d) (R)-3-(tert-butyldimethylsilyloxy)tetradecyl methanesulfonate, NaH, DMF, rt, 16 h, then 50 °C, 1.5 h, 81%; e) p-TsOH, MeOH, rt, 1 h; 50%; f) Ir[(C_8H_{12})(PMePh₂)₂]PF₆ activated by H₂, THF, rt, 16 h, 100%; g) t-BuMe₂SiCl, DMAP, CH₂Cl₂, 75%; h) (1) triphosgene, pyridine, toluene, 0 °C, 10 min, then allyl alcohol, 0 °C, 1 h, 100%; i) aq 48% HF, CH₂Cl₂-MeCN (1:2), rt, 30 min, 67%.

pounds (16α , 16β , 28α , 28β , 40α , and 40β) and their LPS-antagonistic activities toward human whole blood cells, ⁸ C3H/HeN mice⁹ and mouse peritoneal resident macrophages. ¹⁰

Synthesis. Firstly, compounds 16α and 16β were synthesized from both compound 10 obtained from diacetone D-glucose 1 and the known compound 11, reported by an Eisai group. The diacetone D-glucose 1 was alkylated with n-dodecyl methanesulfonate in N, N-dimethylformamide (DMF) using sodium hydride as a base to give dodecyl ether 2. The furanoside 2 was converted to allyl D-pyranoside 3 as a 10:1 anomeric mixture by heating in allyl alcohol containing hydrochloric acid. The separated α -anomer 3α chromatographically was used for the synthesis of 10 (Scheme 1). The 4, 6-diol part of 3α was protected with 2, 2-dimethoxypropane using p-toluenesulfonic acid as a catalyst to yield isopropylidene compound 4. The C-2 free alcohol of 4 was alkylated with (R)-3-(tert-butyl-dimethylsilyloxy)tetradecyl methanesulfonate in DMF using sodium hydride as a base to afford 5. The reagent (R)-3-

(tert-butyldimethylsilyloxy)tetradecyl methanesulfonate was obtained from methyl (R)-3-hydroxytetradecanoate in three steps: 1) silylation of C-3 alcohol with tert-butyldimethylsilyl chloride and imidazole, 2) reduction of methyl ester with LiBH₄-LiEt₃BH, and finally 3) mesylation of the resulting C-1 alcohol by methanesulfonyl chloride and triethylamine in methylene chloride. Both 4,6-O-isopropylidene and the tert-butyldimethylsilyl group on the C-2 side chain were deprotected by a treatment with p-toluenesulfonic acid in methanol to afford triol 6. At this stage, the anomeric allyl compound 6 was converted to vinyl ether 7 by a treatment with (1,5-cyclooctadiene)bis(methyldiphenylphosphine)iridium(I) hexafluorophosphate (Ir[(C₈H₁₂)(PMePh₂)₂]PF₆) activated by hydrogen in order to distinguish it from the allyloxycarbonyl groups introduced as protecting groups in the latter stage. The primary C-6 alcohol of 7 was reprotected with the tert-butyldimethylsilyl group to afford 8. A treatment of the secondary alcohols of 8 with triphosgene using pyridine as a base,

Scheme 2. Reagents and conditions: a) **10**, TMSOTf, AgOTf, MS 4A, CH₂Cl₂, rt, 16 h; b) 1) Zn, AcOH–H₂O (3:2), rt, 3 h, 2) Z-11-octadecenoyl chloride, NaHCO₃, THF–H₂O (3:1), rt, 2 h, three steps 45%; c) aq 48% HF, CH₂Cl₂–CH₃CN (1:2), rt, 16 h, 67%; d) *i*-Pr₂NP(OCH₂CH=CH₂)₂, 1*H*-tetrazole, Na₂SO₄, CH₂Cl₂, rt, 30 min, then aq 30% H₂O₂, THF, 0 °C, 45 min, 49% (**15**α), 50% (**15**β); e) (PPh₃)₄Pd, PPh₃, Et₃N–HCOOH, THF, under N₂, 55 °C, 16 h, 40% (**16**α), 37% (**16**β).

and then allyl alcohol, afforded di-alloc compound **9**, which was further converted to free C-6 alcohol **10** using aqueous 48% hydrofluoric acid (aq 48% HF).

The treatment of alcohol 10 and imidate 11 with trimethylsilyl trifluoromethanesulfonate and silver trifluoromethanesulfonate in methylene chloride gave $\beta(1-6)$ disaccharide 12. The deprotection of the trichloroethoxycarbonyl group from the C-2' trichloroethoxycarbonylamino group of 12 with Zn-acetic acid and the successive acylation of the resulting amine with (Z)-11-octadecenoyl chloride-aq NaHCO₃ in tetrahydrofuran (THF)-water gave amide 13. The treatment of 13 with aq 48% HF afforded an anomeric mixture 14, which was further converted to an anomeric mixture of phosphates (15α) and **15** β) through phosphite by the treatment of diallyl diisopropylphosphoramidite and 1H-tetrazole, and then aqueous 30% hydrogen peroxide. Both α - and β -anomers, 15α and 15β , were separated chromatographically. The treatment of each anomer, 15 α and 15 β , with tetrakis(triphenylphosphine)palladium(0), triphenylphosphine, and triethylamine-formic acid in THF at 55 °C for 16 hours gave 16α and 16β , respectively. Compound **16\beta** was gradually decomposed during long-term storage in a freezer (Scheme 2).

Secondly, compounds 28α and 28β were synthesized from both compound 24 obtained from diacetone D-glucose 1 and the known compound 11 reported by an Eisai group. ⁷ The di-

acetone D-glucose 1 was alkylated with n-decyl methanesulfonate in DMF using sodium hydride as a base to give decyl ether 17. The furanoside 17 was converted to allyl 4,6-O-isopropylidene-D-pyranoside 18 as a 3:1 anomeric mixture by refluxing in allyl alcohol containing hydrochloric acid, and a successive treatment with 2,2-dimethoxypropane using p-toluenesulfonic acid as a catalyst. The C-2 free alcohol of an anomeric mixture 18 was acylated with (R)-3-(tert-butyldimethylsilyloxy)tetradecanoic acid in CH₂Cl₂ using 1-[3-(dimethylamino)propyl]-3-ethylcarbodiimide as a dehydration reagent to give an anomeric mixture 19. The 4,6-O-isopropylidene of 19 was deprotected by a treatment with p-toluenesulfonic acid in methanol to afford an anomeric mixture of diol 20. At this stage, the anomeric allyl compound 20 was converted to an anomeric mixture of vinyl ether 21 by a treatment with Ir[(C₈H₁₂)(PMePh₂)₂]PF₆ activated by hydrogen in order to distinguish it from the allyloxycarbonyl groups introduced as protecting groups in a latter stage. The primary C-6 alcohol of 21 was reprotected with the tert-butyldimethylsilyl group to convert to an anomeric mixture 22. A treatment of the secondary alcohol of 22 with triphosgene and allyl alcohol using pyridine as a base afforded an anomeric mixture of alloc compound 23, which was further converted to an anomeric mixture of free C-1 and C-6 alcohol 24 by treating with aq 48% HF at room temperature for 1.5 days. (Scheme 3).

Scheme 3. Reagents and conditions: a) decyl methanesulfonate, NaH, DMF, rt, 16 h, 98%; b) allyl alcohol containing 2% HCl, reflux, 25 min, 80%; c) 3-oxotetradecanoic acid, 1-[3-(dimethylamino)propyl]-3-ethylcarbodiimide, HCl, CH₂Cl₂, rt, 1 h, 80%; d) p-TsOH•H₂O, MeOH, rt, 3 h, 59%; e) Ir[(C₈H₁₂)(PMePh₂)₂]PF₆ activated by H₂, THF, 16 h, 100%; f) 'BuMe₂SiCl, DMAP, CH₂Cl₂, 16 h, 100%; g) triphosgene, pyridine, toluene, 0 °C, 15 min, then CH₂=CHCH₂OH, 0 °C, 1 h, 68%; h) aq 48% HF, MeCN-CH₂Cl₂ (2:1), rt, 1.5 days, 58%.

The treatment of alcohol 24 and imidate 11 with trimethylsilvl trifluoromethanesulfonate and silver trifluoromethanesulfonate in methylene chloride gave an anomeric mixture on the reducing end of $\beta(1-6)$ disaccharide 25 without reacting with the free anomeric position of 24. Deprotection of the trichloroethoxycarbonyl group of the C-2' trichloroethoxycarbonylamino group of 25 with Zn-acetic acid and successive acylation of the resulting amine with (Z)-11-octadecenoyl chloride-aq NaHCO3 in THF-water gave amide 26 without reacting with the free anomeric position of 25. The treatment of 26 with diallyl diisopropylphosphoramidite and 1H-tetrazole, and then ag 30% hydrogen peroxide afforded an anomeric mixture of phosphates 27α and 27β . The mixture of 27α and 27β was separated chromatographically. The treatment of each anomer, 27α and 27β, with (PPh₃)₄Pd, PPh₃, and Et₃N-HCOOH in THF at 55 °C for 16 hours gave 28α and 28β , respectively (Scheme 4).

Thirdly, compounds 40α and 40β were synthesized from both compound 36, obtained from allyl α -D-glucoside 29, and the known compound 11, reported by an Eisai group. The allyl α -D-glucoside **29**¹¹ was converted to allyl 4,6-O-isopropylidene- α -D-pyranoside 30 by reacting with 2,2-dimethoxypropane using p-toluenesulfonic acid as a catalyst. The C-2 and C-3 free alcohols of 30 were alkylated with dodecyl iodide and NaH as a base to afford 31. The 4,6-O-isopropylidene of 31 was deprotected by a treatment with p-toluenesulfonic acid in methanol to afford diol 32. At this stage, the allyl compound 32 was converted to vinyl ether 33 by a treatment with Ir[(C₈H₁₂)(PMePh₂)₂]PF₆ activated by hydrogen in order to distinguish it from the allyloxycarbonyl groups introduced as protecting groups in a latter stage. The primary C-6 alcohol of 33 was reprotected to be *tert*-butyldimethylsilyl ether 34. A treatment of the secondary alcohol of 34 with triphosgene and allyl alcohol using pyridine as a base afforded allyloxycarbonyl compound **35**, which was further converted to an anomeric mixture of free C-1 and C-6 alcohols **36** by treating with aq 48% HF at room temperature for 16 hours. (Scheme 5).

The treatment of alcohol 36 and imidate 11 with trimethylsilvl trifluoromethanesulfonate and silver trifluoromethanesulfonate in methylene chloride gave an anomeric mixture of $\beta(1-6)$ disaccharide 37 without reacting with the free anomeric position of 36. Deprotection of the C-2 trichloroethoxycarbonyl group of 37 with Zn-acetic acid, and successive acylation of the resulting amine with (Z)-11-octadecenoyl chloride-aqueous NaHCO3 in THF-water gave amide 38 without reacting with the free anomeric position of 37. The treatment of 38 with diallyl diisopropylphosphoramidite and 1H-tetrazole, and then aq 30% hydrogen peroxide converted to an anomeric mixture of phosphates 39α and 39β . The mixture of 39α and 39β was separated chromatographically. The treatment of each anomer, 39α and 39β , with (PPh₃)₄Pd, PPh₃, and Et₃N-HCOOH in THF at 55 °C for 16 hours gave 40α and **40** β , respectively (Scheme 6).

Thus, we could synthesize six disaccharides $(16\alpha, 16\beta, 28\alpha, 28\beta, 40\alpha, \text{ and } 40\beta)$.

Biological Activity. The inhibitory activity on LPS-induced TNF α production, LPS-antagonistic activity, of three pairs of synthetic compounds (16 α and 16 β , 28 α and 28 β , and 40 α and 40 β) was investigated in vitro using human whole blood cells. The IC₅₀ values (nM) of these six compounds (16 α , 16 β , 28 α , 28 β , 40 α , and 40 β) toward human whole blood cells were 0.35, 0.42, 2.37, 1.16, 2.89, and 7.70, respectively. The activities of these compounds were generally strong, except for 40 β . The activity of the unknown type β -anomer 40 β in nature was almost one-third compared with its corresponding α -anomer 40 α . However, astonishingly, the activity of β -anomer 28 β was two-times stronger than that of natural type 28 α , and 16 α and 16 β had almost the same lev-

Scheme 4. Reagents and conditions: a) **24**, TMSOTf, AgOTf, MS 4A, CH₂Cl₂, 24 °C, 97%; b) 1) Zn, AcOH–THF (1:1), rt, 3 h; 2) (Z)-11-octadecenoyl chloride, NaHCO₃, THF–H₂O (3:1), rt, 2 h, 55%; c) *i*-Pr₂NP(OCH₂CH=CH₂)₂, 1*H*-tetrazole, Na₂SO₄, CH₂Cl₂, rt, 30 min, then aq 30% H₂O₂, THF, rt, 30 min, 22% (**27**α), 20% (**27**β); d) (PPh₃)₄Pd, PPh₃, Et₃N–HCOOH, THF, under N₂, 55 °C, 16 h, 59% (**28**α), 72% (**28**β).

Scheme 5. Reagents and conditions: a) 2,2-dimethoxypropane, *p*-TsOH, DMF, rt, 16 h, 65%; b) dodecyl iodide, NaH, DMF, rt, 16 h, 98%; c) *p*-TsOH, MeOH, rt, 30 min, 54%; d) Ir[(C₈H₁₂)(PMePh₂)₂]PF₆ activated by H₂, THF, N₂, rt, 16 h, 100%; e) *t*-BuMe₂SiCl, DMAP, CH₂Cl₂, 95%; f) triphosgene, pyridine, toluene, 0 °C, 10 min, then allyl alcohol, 0 °C, 1 h, 100%; g) aq 48% HF, CH₂Cl₂-MeCN (1:2), rt, 16 h, 74%.

el of activity.

Also, the inhibitory activity on TNF α production toward galactosamine loaded C3H/HeN mice of compounds 28α and 28β was measured, and the activities were sufficiently strong. The values of these compounds were 1.05 and 3.10 nM, respectively. The natural type α -anomer was three-times stronger than the unknown type β -anomer in nature. Thus, the LPS-antagonistic activity toward human whole blood and

the inhibitory activity on TNF α production toward galactosamine loaded C3H/HeN mice of compounds 28α and 28β were not in parallel.

Also, the inhibitory activities (IC₅₀) on LPS-induced TNF α production of a pair of synthetic compounds, 40α and 40β , toward mouse peritoneal resident macrophages were 64 and 58 nM, respectively. The configuration of the anomeric position on the reducing glucose had no influence on the activity.

Scheme 6. Reagents and conditions: a) **36**, TMSOTf, AgOTf, MS 4A, CH_2Cl_2 , rt, 16 h, 96%; b) 1) Zn, AcOH–THF (1:1), rt, 2.5 h, 2) (*Z*)-11-octadecenoyl chloride, NaHCO₃, THF–H₂O (3:1), rt, 1 h, 65%; c) *i*-Pr₂NP(OCH₂CH=CH₂)₂, 1*H*-tetrazole, Na₂SO₄, CH_2Cl_2 , rt, 30 min, then aq 30% H_2O_2 , THF, 0 °C, 20 min, 25% (**39** α), 26% (**39** β); d) (PPh₃)₄Pd, PPh₃, Et₃N–HCOOH, THF, under N₂, 55 °C, 16 h, 54% (**40** α), 52% (**40** β).

As a result, even for un-natural type β -anomers, it may be true that they are still sufficiently active, or even stronger than the corresponding α -anomers in LPS-antagonistic activity toward human whole blood cells, C3H/HeN mice and mouse peritoneal resident macrophages.¹²

Usually, lipid A analogs having six fatty acid chains show LPS-agonistic (endotoxic) activity toward both human U-937 and mouse peritoneal resident macrophages, and lipid IVa¹³ having four fatty acid chains shows LPS-antagonistic activity toward human blood cells, and adversely endotoxic activity toward mouse peritoneal resident macrophages. This fact shows, interestingly enough, that a difference exists in molecular recognition between human and mouse LPS receptors. ¹⁴ The synthetic compounds (16α , 16β , 28α , 28β , 40α , and 40β), this time, showed LPS-antagonistic activity toward both human and mouse blood cells, as anticipated from E5564. This tendency was the same activity as that for a nontoxic natural RsDPLA⁵ having a cis-double bond in one of the fatty acid chains isolated from *Rhodobacter sphaeroides*.

Experimental

 1 H NMR spectra were recorded with JEOL-GSX 400 and JNM-ECT 500 spectrometers using tetramethylsilane (TMS) as an internal standard. 13 C NMR spectra were recorded on a JNM-ECT (500 MHz) at 125 MHz. IR absorption spectra were measured with an IR A-2 spectrophotometer, and mass spectra were obtained with a JMS-700 mass spectrometer. The separation of compounds by column chromatography was done with silica-gel 60 (230–400 mesh ASTM) under a slightly elevated pressure (111–182 kPa) for easy elution. Analytical chromatography was performed on Merck Art 5715 silica-gel 60- F_{245} plates. Commercially available anhydrous THF and dichloromethane were used for the reactions. DMF and pyridine were dried by storage over 4 Å molecular sieves.

(R)-3-(tert-Butyldimethylsilyloxy)tetradecyl methanesulfonate. (a) To a solution of (R)-methyl 3-hydroxytetradecanoate (10.00 g, 38.70 mmol) in DMF (25 mL) were added tert-butyldimethylsilyl chloride (9.80 g, 58.05 mmol) and imidazole (6.50 g, 95.48 mmol). The mixture was stirred for 16 h at room temperature, and diluted with EtOAc, washed with brine, dried over MgSO₄, and filtered. The filtrate was concentrated in vacuo to give a residue, which was chromatographed on a silica-gel column. Elution with cyclohexane–EtOAc (19:1) gave (R)-methyl 3-(tert-butyldimethylsilyloxy)tetradecanoate (14.3 g, 99%) as an oil

(b) To a suspension of LiBH₄ (1.76 g, 80.8 mmol) in dry Et₂O (150 mL) was added a solution of the above-obtained (R)-methyl 3-(tert-butyldimethylsilyloxy)tetradecanoate (25.00 g, 67.1 mmol) in dry Et₂O (50 mL) at room temperature under N₂. To this solution was added a solution of LiEt₃BH (1 M in THF, 6.56 mL). The mixture was refluxed for 2 h. After cooling, the mixture was poured into sat. aq NH₄Cl (150 mL) at ice cooling temperature, and extracted with ether. The extract was washed with brine, dried over MgSO₄, and filtered. The filtrate was concentrated in vacuo to give a mixture, which was chromatographed on a silica-gel column. Elution with cyclohexane-EtOAc (7:1) gave (R)-3-(tertbutyldimethylsilyloxy)tetradecanol (22.9 g, 99%) as an oil. 400 MHz ¹H NMR (CDCl₃) δ 0.08 (3H, s), 0.09 (3H, s), 0.88 (3H, t, J = 6.6 Hz), 0.90 (9H, s), 1.26 (18H, brs), 1.51–1.59 (2H, m), 1.64 (1H, m), 1.82 (1H, m), 2.49 (1H, brs, OH), 3.72 (1H, m), 3.84 (1H, m), 3.92 (1H, m).

(c) To a solution of the above-obtained (R)-3-(tert-butyldimethylsilyloxy)tetradecanol (1.79 g, 5.19 mmol) and Et_3N (630 mg, 6.23 mmol) in dry CH_2Cl_2 (15 mL) was added dropwise a solution of methanesulfonyl chloride (720 mg, 6.28 mmol) in dry CH_2Cl_2 (5 mL) at 0 °C under nitrogen. After stirring for 15 min, the mixture was diluted with CH_2Cl_2 , and washed with H_2O , sat. aq $NaHCO_3$, and brine, dried over $MgSO_4$, and filtered.

The filtrate was concentrated to give the title compound quantitatively. 400 MHz 1 H NMR (CDCl₃) δ 0.06 (6H, s), 0.89 (9H, s), 0.89 (3H, t, J=6.6 Hz), 1.26 (18H, brs), 1.45–1.48 (2H, m), 1.85 (1H, m), 1.92 (1H, m), 3.00 (3H, s), 3.82 (1H, m), 4.29–4.35 (2H, m). Anal. Calcd for C₂₁H₄₆O₄SSi: C, 59.66; H, 10.97; S, 7.59%. Found: C, 59.53; H, 10.61; S, 7.85%.

1,2:5,6-Di-*O***-isopropylidene-3-***O***-dodecyl-**α-**D-glucofuranose** (2). To a solution of **1** (13.02 g, 50.0 mmol) in DMF (50 mL) and dodecyl methanesulfonate (13.20 g, 50.0 mmol) was added NaH (55% oil dispersion, 2.88 g, 60.0 mmol) at 0 °C. After stirring for 15 min, the mixture was stirred for 16 h at room temperature, quenched with MeOH, diluted with EtOAc, which was washed with water and brine, dried over MgSO₄, and filtered. The filtrate was concentrated to give a mixture, which was chromatographed on a silica-gel column. Elution with cyclohexane–EtOAc (9:1) gave **2** (15.50 g, 75%) as an oil. 400 MHz ¹H NMR (CDCl₃) δ 0.88 (3H, t, J = 6.6 Hz), 1.26 (18H, brs), 1.32 (3H, s), 1.35 (3H, s), 1.43 (3H, s), 1.50 (3H, s), 1.52–1.57 (2H, m), 3.51 (1H, m), 3.59 (1H, m), 3.85 (1H, d, J = 2.9 Hz), 3.98 (1H, m), 4.06–4.14 (3H, m), 4.31 (1H, m), 4.52 (1H, d, J = 3.7 Hz), 5.88 (1H, d, J = 3.7 Hz).

Allyl 3-*O*-dodecyl- α , β -D-glucopyranoside (3 α , 3 β). A solution of 2 (900 mg, 2.160 mmol) in allyl alcohol (10 mL) containing 2% HCl was refluxed for 20 min. The reaction mixture was concentrated in vacuo to give a mixture, which was chromatographed on a silica-gel column. Elution with cyclohexane-EtOAc (1:1, then 1:3) gave 3α (674 mg, 80%) as a solid and 3β (64 mg, 8%) as an oil. Physical data of 3α : $R_f = 0.230$ (cyclohexane– EtOAc (1:1)). mp 72–73.5 °C (needles). IR ν_{max} (KBr) 3306, 2920, 2852 cm⁻¹. 400 MHz ¹H NMR (CDCl₃) δ 0.88 (3H, t, J =6.6 Hz), 1.26 (18H, brs), 1.59–1.64 (2H, m), 2.19 (1H, d, J = 9.5Hz, OH), 2.24 (1H, t, J = 6.3 Hz, OH), 2.79 (1H, d, J = 2.9 Hz, OH), 3.45 (1H, d, J = 8.8 Hz), 3.50-3.61 (2H, m), 3.64-3.71 (2H, m), 3.80–3.86 (2H, m), 3.93 (1H, m), 4.04 (1H, m), 4.23 (1H, m), 4.90 (1H, d, J = 3.7 Hz), 5.22-5.34 (2H, m), 5.92 (1H, m). Anal.Calcd for C₂₁H₄₀O₆: C, 64.92; H, 10.38%. Found: C, 64.96; H, 10.17%. Physical data of 3β : $R_f = 0.308$ (cyclohexane–EtOAc (1:1)). 400 MHz ¹H NMR (CDCl₃) δ 0.88 (3H, t, J = 7.0 Hz), 1.26 (18H, brs), 1.50-1.61 (2H, m), 3.46-3.53 (10H, m), 5.07 (1H, d, J = 5.1 Hz), 5.21-5.42 (2H, m), 5.93 (1H, m).

Allyl 3-*O*-dodecyl-4,6-*O*-isopropylidene-α-D-glucopyranoside (4). A solution of 3α (470 mg, 1.210 mmol) in DMF (1 mL) and 2,2-dimethoxypropane (1 mL) containing *p*-TsOH·H₂O (20 mg) was stirred for 16 h at room temperature. The reaction mixture was concentrated in vacuo to give a mixture, which was chromatographed on a silica-gel column. Elution with cyclohexane–EtOAc (4:1, then 2:1) gave 4 (420 mg, 81%) as an oil. 400 MHz ¹H NMR (CDCl₃) δ 0.88 (3H, t, J = 6.6 Hz), 1.26 (18H, brs), 1.41 (3H, s), 1.49 (3H, s), 1.53–1.60 (2H, m), 2.31 (1H, d, J = 7.3 Hz, OH), 3.46–3.85 (8H, m), 4.04 (1H, m), 4.21 (1H, m), 4.92 (1H, d, J = 3.7 Hz), 5.22–5.34 (2H, m), 5.93 (1H, m).

Allyl 2-O-[(R)-3-(tert-butyldimethylsilyloxy)tetradecyl]-3-O-dodecyl-4,6-O-isopropylidene-α-D-glucopyranoside (5). To a solution of 4 (4.91 g, 11.45 mmol) in DMF (30 mL) were added (R)-3-(tert-butyldimethylsilyloxy)dodecyl methanesulfonate (5.37 g, 12.71 mmol) and NaH (55% oil dispersion, 1.50 g, 34.38 mmol). The mixture was stirred for 16 h at room temperature, and then for 1.5 h at 50 °C, and diluted with EtOAc, which was washed with ice water and brine, dried over MgSO₄, and filtered. The filtrate was concentrated in vacuo to give a residue, which was chromatographed on a silica-gel column. Elution with cyclohexane–EtOAc (9:1) gave 5 (7.01 g, 81%) as an oil. IR ν_{max}

(film) 2925, 2856, 1464 cm⁻¹. 400 MHz ¹H NMR (CDCl₃) δ 0.04 (6H, s), 0.86–0.89 (15H, m, containing 9H, s, at δ 0.88 ppm), 1.26 (38H, brs), 1.40 (3H, s), 1.48 (3H, s), 1.48–1.55 (2H, m), 1.70–1.77 (2H, m), 3.29 (1H, m), 3.45–3.78 (9H, m), 3.84 (1H, m), 4.04 (1H, m), 4.17 (1H, m), 4.91 (1H, d, J=3.7 Hz), 5.20–5.34 (2H, m), 5.92 (1H, m). FABMS (positive-ion) m/z 755 [M+H]⁺, 777 [M+Na]⁺ (on addition of NaI). HRFABMS Calcd for $C_{44}H_{86}O_7SiNa$: 777.6040. Found: 777.5994.

Allyl 3-O-dodecyl-2-O-[(R)-3-hydroxytetradecyl]-α-D-glucopyranoside (6). To a solution of 5 (965 mg, 1.287 mmol) in MeOH (20 mL) was added TsOH·H₂O (50 mg, 0.263 mmol). The mixture was stirred for 40 min at room temperature, and diluted with EtOAc, which was washed with sat. aq NaHCO3 and brine, dried over MgSO₄, and filtered. The filtrate was concentrated in vacuo to give a residue, which was chromatographed on a silica-gel column. Elution with cyclohexane-EtOAc (1:1), and then EtOAc gave 6 (386 mg, 50%) as a solid. IR ν_{max} (KBr) 3400-3260, 2920, 2850 cm⁻¹. 400 MHz ¹H NMR (CDCl₃) 0.86-0.89 (6H, m), 1.26 (36H, brs), 1.41-1.73 (6H, m), 2.07 (1H, brs, OH), 2.60 (1H, brs, OH), 2.96 (1H, brs, OH), 3.30 (1H, m), 3.50-3.89 (10H, m), 4.04 (1H, m), 4.20 (1H, m), 5.00 (1H, m), 5.22-5.34 (2H, m), 5.95 (1H, m). FABMS (positiveion) m/z 599 $[M-H]^+$, 601 $[M+H]^+$, 623 $[M+Na]^+$. HRFABMS Calcd for C₃₅H₆₈O₇Na: 623.4862. Found: 623.4861.

(E)-1-Propenyl 3-O-dodecyl-2-O-[(R)-3-hydroxytetradecyl]- α -D-glucopyranoside (7). To a solution of 6 (381 mg, 0.634 mmol) in dry THF (5 mL) was added (1,5-cyclooctadiene)bis-(methyldiphenylphosphine)iridium(I) hexafluorophosphate mg). The atmosphere of the flask bottle was replaced with nitrogen, and then hydrogen to activate the iridium complex. After the red iridium complex turned colorless in solution upon stirring for about 10 seconds under a H2 atmosphere, the hydrogen was replaced again by nitrogen. The solution was stirred for 16 h at room temperature, and concentrated in vacuo to give 7 (381 mg, 100%) as a solid. IR ν_{max} (KBr) 3411 (br), 2920, 2850, 1679 (w), 1468 cm⁻¹. 400 MHz ¹H NMR (CDCl₃) δ 0.88 (6H, t, J =6.6 Hz), 1.26 (36H, brs), 1.41-1.73 (9H, m), 1.86 (1H, brs, OH), 2.45 (1H, brs, OH), 2.72 (1H, brs, OH), 3.32 (1H, dd, J = 3.7, 9.5 Hz), 3.49–3.92 (10H, m), 5.16–5.23 (2H, m), 6.19 (1H, dd, J = 1.5, 12.5 Hz). FABMS (positive-ion) m/z 601 $[M + H]^+$, 623 $[M + Na]^+$. HRFABMS Calcd for $C_{35}H_{68}O_7Na$: 623.4862. Found: 623.4880.

(*E*)-1-Propenyl 6-*O-tert*-butyldimethylsilyl-3-*O*-dodecyl-2-*O*-[(*R*)-3-hydroxytetradecyl]-α-D-glucopyranoside (8). To a solution of 7 (747 mg, 1.243 mmol) in CH₂Cl₂ (15 mL) were added *tert*-BuMe₂SiCl (225 mg, 1.492 mmol) and DMAP (182 mg, 1.492 mmol). The solution was stirred for 16 h at room temperature. The mixture was directly chromatographed on a silica-gel column. Elution with cyclohexane–EtOAc (9:1, and then 2:1) gave 8 (670 mg, 75%) as a powder. IR $\nu_{\rm max}$ (KBr) 3226, 2925, 2853, 1749, 1467 cm⁻¹. 400 MHz ¹HNMR (CDCl₃) δ 0.08 (6H, s), 0.88 (3H, t, J = 6.6 Hz), 0.90 (9H, s), 1.26 (36H, brs), 1.40–1.73 (7H, m, containing 3H, dd, J = 1.5, 6.6 Hz, at δ 1.55 ppm), 2.74 (1H, brs, OH), 2.81 (1H, brs, OH), 3.30 (1H, m), 3.51–3.87 (10H, m), 5.14 (1H, d, J = 3.7 Hz). FABMS (positive-ion) m/z 715 [M + H]⁺, 737 [M + Na]⁺. HRFABMS Calcd for C₄₁H₈₂O₇SiNa: 737.5728. Found: 737.5730.

(*E*)-1-Propenyl 4-*O*-allyloxycarbonyl-6-*O*-tert-butyldimethylsilyl-3-*O*-dodecyl-2-*O*-[(R)-3-(allyloxycarbonyloxy)tetradecyl]- α -D-glucopyranoside (9). To a solution of **8** (659 mg, 0.921 mmol) in toluene (20 mL) and pyridine (1.57 g) was added triphosgene (623 mg, 2.099 mmol) at 0 °C. After stirring for 10

min, allyl alcohol (1.88 g) was added to this solution. The mixture was stirred for 1 h at 0 °C, and diluted with EtOAc, which was washed with sat. aq NaHCO₃ and brine, dried over MgSO₄, and filtered. The filtrate was concentrated in vacuo to give a residue, which was chromatographed on a silica-gel column. Elution with cyclohexane-EtOAc (14:1) gave 9 (813 mg, 100%) as an oil. IR ν_{max} (film) 2927, 2856, 1749, 1680 (w), 1464, 1371 cm⁻¹. 400 MHz 1 H NMR (CDCl₃) δ 0.02 (3H, s), 0.03 (3H, s), 0.86–0.90 (15H, m, containing 9H, s, at δ 0.88 ppm), 1.25 (36H, brs), 1.46–1.63 (7H, m, containing 3H, dd, J = 2.2, 6.6 Hz, at δ 1.54 ppm), 1.88–1.91 (2H, m), 3.35 (1H, m), 3.53 (1H, m), 3.65– 3.80 (7H, m), 4.59–4.65 (4H, m), 4.71 (1H, t, J = 9.9 Hz), 4.78 (1H, m), 5.09 (1H, d, J = 3.7 Hz), 5.18 (1H, m), 5.25–5.39 (4H, m), 5.89–5.98 (2H, m), 6.17 (1H, qd, J = 2.2, 12.5 Hz). FABMS (positive-ion) m/z 883 $[M + H]^+$, 905 $[M + Na]^+$. HRFABMS Calcd for C₄₉H₉₀O₁₁SiNa: 905.6150. Found: 905.6188.

(E)-1-Propenyl 4-O-allyloxycarbonyl-2-O-[(R)-3-(allyloxycarbonyloxy)tetradecyl]-3-O-dodecyl-α-D-glucopyranoside (10). To a solution of 9 (840 mg, 0.951 mmol) in CH₂Cl₂ (5 mL) and MeCN (10 mL) were added silica-gel powder (150 mg) and aq 48% HF (100 mg). The mixture was stirred for 13 min at room temperature, and diluted with EtOAc, which was washed with sat. aq NaHCO3 and brine, dried over MgSO4, and filtered. The filtrate was concentrated in vacuo to give a residue, which was chromatographed on a silica-gel column. Elution with cyclohexane-EtOAc (2:1) gave 10 (485 mg, 67%) as an oil. IR ν_{max} (film) 3527 (br), 2925, 2854, 1747, 1679 (w) cm⁻¹. 400 MHz ¹H NMR $(CDCl_3)$ 0.88 (6H, t, J = 6.6 Hz), 1.25 (36H, brs), 1.50–1.65 (7H, m, containing 3H, dd, J = 1.5, 6.6 Hz, at δ 1.55 ppm), 1.87–1.92 (2H, m), 2.33 (1H, brs, OH), 3.37 (1H, dd, J = 2.9, 9.5 Hz), 3.55– 3.81 (8H, m), 4.60–4.67 (4H, m), 4.72 (1H, m), 4.80 (1H, m), 5.13 (1H, d, J = 3.7 Hz), 5.18 (1H, qd, J = 6.6, 12.5 Hz), 5.25-5.40(4H, m), 5.89–5.98 (2H, m), 6.16 (1H, qd, J = 1.5, 12.5 Hz). FABMS (positive-ion) m/z 768 $[M + H]^+$, 791 $[M + Na]^+$. HRFABMS Calcd for C₄₃H₇₆O₁₁Na: 791.5286. Found: 791.5281.

(E)-1-Propenyl 4-O-allyloxycarbonyl-2-O-[(R)-3-(allyloxycarbonyloxy)tetradecyl]-6-O-{2-deoxy-4-O-diallylphosphono-3-O-[(R)-3-(methoxy)decyl]-6-O-methyl-2-[(trichloroethoxycarbonyl)amino]- β -D-glucopyranosyl}-3-O-dodecyl- α -D-glucopyranoside (12). A solution of 10 (460 mg, 0.598 mmol) and imidate 11 (606 mg, 0.718 mmol) in CH₂Cl₂ (17 mL) cotaining MS 4A (1.4 g) was stirred for 20 min at room temperature under nitrogen. To this mixture were added AgOTf (310 mg, 1.207 mmol) and TMSOTf (27 mg, 0.121 mmol). This mixture was stirred for 16 h at room temperature under nitrogen, and diluted with CH₂Cl₂, which was washed with sat. aq NaHCO₃ and brine, dried over MgSO₄, and filtered. The filtrate was concentrated in vacuo to give a residue, which was chromatographed on a silica-gel column. Elution with cyclohexane-EtOAc (2:1) gave 12 (867 mg, 100%) as a gum. IR ν_{max} (film) 3400–3080 (w), 2927, 2856, 1745 cm⁻¹. 400 MHz ¹H NMR (CDCl₃) δ 0.88 (9H, t, J = 6.6Hz), 1.25-1.30 (48H, m), 1.38-1.75 (9H, m), 1.85-1.90 (2H, m), 3.23–3.80 (22H, m, containing two 3H, s, at δ 3.28 and 3.38 ppm), 4.04 (1H, d, J = 11.7 Hz), 4.26 (1H, dd, J = 9.5, 18.3 Hz), 4.51–4.85 (12H, m), 5.10 (1H, d, J = 3.7 Hz), 5.16 (1H, m), 5.23–5.40 (8H, m), 5.89–5.99 (4H, m), 6.16 (1H, dd, J = 1.5, 12.5 Hz). FABMS (positive-ion) m/z 1470 [M + Na,³⁵Cl]⁺, 1472. HRFABMS Calcd for C₇₀H₁₂₁Cl₃NO₂₁PNa: 1470.7132. Found: 1470.7102.

 $\label{eq:continuous} \begin{array}{ll} (E)\text{-1-Propenyl} & 4\text{-}O\text{-allyloxycarbonyl-3-}O\text{-dodecyl-6-}O\text{-}\{2\text{-deoxy-4-}O\text{-diallylphosphono-3-}O\text{-}[(R)\text{-3-(methoxy)decyl]-6-}O\text{-}(R)\text{-3-(methoxy)decyl]-6-}O\text{-}(R) \end{array}$

methyl-2-[(Z)-11-(octadecenoyl)amino]- β -D-glucopyranosyl}-2-O-[(R)-3-(allyloxycarbonyloxy)tetradecyl]- α -D-glucopyranoside (13). To a solution of 12 (895 mg, 0.617 mmol) in THF-AcOH (3:2, 35 mL) was added Zn powder (800 mg, 12.23 m atom). The mixture was stirred vigorously with a magnetic stirrer at 25 °C for 3 h, and filtered. The filtrate was concentrated in vacuo below 30 °C, and diluted with EtOAc, which was washed with sat. NaHCO₃, and brine, dried over MgSO₄, and filtered. The filtrate was concentrated in vacuo to give amine, which was dissolved in THF (6 mL)-H₂O (2 mL) containing NaHCO₃ (220 mg, 2.619 mmol). To this solution was added a solution of cisvaccenyl chloride (250 mg, 0.831 mmol) in THF (4 mL) with vigorous stirring at 25 °C. (This acid chloride was prepared from (Z)-11-octadecenoic acid and excess oxalyl chloride in benzene at room temperature for 3 h.) After stirring for 2 h, the reaction mixture was diluted with EtOAc, which was washed with sat. aq NaHCO3 and brine, dried over MgSO4, filtered. The filtrate was concentrated in vacuo to give a mixture, which was chromatographed on a silica-gel column. Elution with cyclohexane-EtOAc (3:1) gave **13** (428 mg, 45%) as an oil. IR ν_{max} (film) 3306 (w), 2926, 2855, 1747, 1678, 1659, 1547, 1465 cm⁻¹. 400 MHz ¹H NMR (CDCl₃) 0.88 (12H, t, J = 6.6 Hz), 1.25 (66H, brs), 1.40–1.80 (13H, m, containing 3H, dd, J = 1.5, 6.6 Hz, at δ 1.54 ppm), 1.84-1.89 (2H, m), 2.00-2.05 (4H, m), 2.22-2.26 (2H, m), 3.22–3.79 (20H, m, containing two 3H, s, at δ 3.28 and 3.37 ppm), 3.85 (1H, m), 4.04 (1H, m), 4.24 (1H, m), 4.53-4.72 (10H, m), 4.78–4.84 (2H, m), 5.09 (1H, d, J = 3.7 Hz), 5.16 (1H, m), 5.22-5.41 (10H, m), 5.88-5.99 (4H, m), 6.05 (1H, d, J = 8.1 Hz), 6.13 (1H, dd, J = 1.5, 12.5 Hz). FABMS (positive-ion) m/z 1560 [M + Na]⁺. HRFABMS Calcd for C₈₅H₁₅₂NO₂₀PNa: 1561.0543. Found: 1561.0560.

4-O-Allyloxycarbonyl-2-O-[(R)-3-(allyloxycarbonyloxy)tetradecyl]-3-O-dodecyl-6-O-{2-deoxy-4-O-diallylphosphono-3-O-[(R)-3-(methoxy)decyl]-6-O-methyl-2-[(Z)-11-(octadecenoyl)amino]- β -D-glucopyranosyl}-D-glucopyranose (14). To a solution of 13 (421 mg, 0.274 mmol) in CH₂Cl₂ (7.7 mL) and MeCN (16.8 mL) was added an aq 48% HF solution (5.3 mL). The mixture was stirred vigorously for 16 h at room temperature, and diluted with EtOAc, which was washed with H₂O, sat. aq NaHCO₃ and brine, dried over MgSO₄, and filtered. The filtrate was concentrated in vacuo to give a residue, which was chromatographed on a silica-gel column. Elution with cyclohexane-EtOAc (1:1, then 1:3) gave an anomeric mixture 14 (274 mg, 67%) as a gum. IR ν_{max} (film) 3300 (w), 2926, 2855, 1748, 1652, 1549, 1465 cm⁻¹. 400 MHz ¹H NMR (CDCl₃) δ 0.86–0.88 (12H, m), 1.25 (66H, brs), 1.40-1.90 (13H, m, containing OH), 1.92-2.04 (4H, m), 2.16–2.19 (2H, m), 3.08–3.77 (20H, m, containing 3H, s, at δ 3.27 ppm, and 3H, s, at δ 3.38 ppm), 3.85–4.63 (14H, m), 4.82 (1H, m), 5.23-5.40 (11H, m), 5.88-5.98 (4H, m), 6.05 (1H, d, J = 7.3 Hz, NH). FABMS (positive-ion) m/z1520 $[M + Na]^+$. HRFABMS Calcd for $C_{82}H_{148}NO_{20}PNa$: 1521.0230. Found: 1521.0250.

Diallylphosphono 4-O-allyloxycarbonyl-3-O-dodecyl-6-O-{2-deoxy-4-O-diallylphosphono-3-O-[(R)-3-(methoxy)decyl]-6-O-methyl-2-[(Z)-11-(octadecenoyl)amino]- β -D-glucopyranosyl}-2-O-[(R)-3-(allyloxycarbonyloxy)tetradecyl]- α -D-glucopyranoside (15 α) and Diallylphosphono 4-O-allyloxycarbonyl-3-O-dodecyl-6-O-{2-deoxy-4-O-diallylphosphono-3-O-[(R)-3-(methoxy)decyl]-6-O-methyl-2-[(Z)-11-(octadecenoyl)amino]- β -D-glucopyranosyl}-2-O-[(R)-3-(allyloxycarbonyloxy)tetradecyl]- β -D-glucopyranoside (15 β). To a solution of 14 (265 mg, 0.177 mmol) in CH₂Cl₂ (10 mL) were added Na₂SO₄ (570

mg), 1H-tetrazole (410 mg, 5.853 mmol) and diallyl diisopropylphosphoramidite (450 mg, 1.834 mmol). After stirring for 30 min at room temperature, this reaction mixture was directly charged on a silica-gel column, and elution with cyclohexane-EtOAc (1:1) gave phosphite (304 mg) as an oil. This phosphite was dissolved in THF (25 mL), and an aq 31% H₂O₂ solution (0.60 mL) was added at 0 °C to this solution. The mixture was stirred for 45 min at 0 °C, and diluted with EtOAc, which was washed with aq 10% Na₂S₂O₃, sat. NaHCO₃, and brine, dried over MgSO₄, and filtered. The filtrate was concentrated in vacuo to give a mixture, which was chromatographed on a silica-gel column. Elution with cyclohexane–EtOAc (1:1, then 1:3) gave 15β (147 mg, 50% yield, $R_f = 0.500$ [cyclohexane–EtOAc (2:3)]) and 15α (144 mg, 49%, $R_f = 0.357$ [cyclohexane–EtOAc (2:3)]) as a gum. Physical data of **15** β : IR ν_{max} (film) 3309 (w), 2926, 2856, 1748, 1678, 1550, 1462 cm $^{-1}$. 400 MHz 1 H NMR (CDCl₃) δ 0.86–0.90 (12H, m), 1.26 (66H, brs), 1.40-2.07 (16H, m), 2.21-2.24 (2H, m), 3.18–3.86 (22H, m, containing two 3H, s, at δ 3.27 and 3.38 ppm), 3.99 (1H, m), 4.25 (1H, dd, J = 9.5, 18.3 Hz), 4.43– 4.65 (12H, m), 4.70 (1H, d, J = 8.8 Hz), 4.78 (1H, m), 4.93 (1H, dd, J = 5.1, 8.1 Hz), 5.19–5.43 (14H, m), 5.86–6.00 (6H, m), 7.48 (1H, d, J = 9.5 Hz, NH). FABMS (positive-ion) m/z1680 $[M+Na]^+$. HRFABMS Calcd for $C_{88}H_{157}NO_{23}P_2Na$: 1681.05. Found: 1681.05. Physical data of 15 α : IR ν_{max} (film) 3306 (w), 2926, 2856, 1747, 1660, 1548 cm⁻¹. 400 MHz ¹H NMR (CDCl₃) δ 0.86–0.90 (12H, m), 1.25 (66H, brs), 1.40–2.05 (16H, m), 2.21-2.25 (2H, m), 3.22-4.06 (23H, m, containing two 3H, s, at δ 3.28 and 3.38 ppm), 4.26 (1H, dd, J = 9.5, 18.3 Hz), 4.54– 4.62 (12H, m), 4.64–4.73 (2H, m), 4.78 (1H, t, J = 5.9 Hz), 5.23-5.43 (14H, m), 5.72 (1H, dd, J = 3.7, 6.6 Hz), 5.88-6.01(6H, m), 6.54 (1H, d, J = 8.1 Hz, NH). FABMS (positive-ion) m/z 1680 [M + Na]⁺.

Phosphono $6-O-\{2-\text{deoxy-}3-O-[(R)-3-(\text{methoxy})\text{decyl}]-6-O$ methyl-2-[(Z)-11-(octadecenoyl)amino]-4-O-phosphono- β -Dglucopyranosyl}-3-O-dodecyl-2-O-[(R)-3-(hydroxy)tetradecyl]- α -D-glucopyranoside (16 α). To a solution of 15 α (140 mg, 0.084 mmol) in dry THF (8 mL) were added PPh3 (20 mg, 0.133 mmol), Et₃N (80 mg, 0.791 mmol), HCOOH (73 mg, 1.457 mmol), and Pd(PPh₃)₄ (20 mg, 0.017 mmol) in this sequence. The solution was stirred for 20 h at 55 °C under nitrogen, and concentrated in vacuo to give a mixture, which was chromatographed on a DEAE-cellulose (Whatman Ion-Exchange cellulose, wet 3 g) column. The column was prepared by preliminary consecutive washing with 30 mL each of 0.5 M HCl, H2O, 0.5 M NaOH, and H₂O, and 12 mL each of 1 M AcOH and H₂O, and 30 mL of 0.05 M AcO·NH₄, 30 mL each of 2:3:1 CHCl₃-MeOH-H₂O, and finally 2:1 CHCl₃-MeOH. The column was eluted with 3 mL each of 2:1 CHCl3-MeOH, then 0.05 M AcO·NH₄ in 2:3:1 CHCl₃-MeOH-H₂O. The fractions containing 16α were collected. To this solution were added another volume of CHCl₃ and aq 0.15 M HCl to adjust the ratio of CHCl₃-H₂O-MeOH to 1:1:1, and the mixture was shaken well. The lower CHCl₂ layer was separated, and concentrated in vacuo to give 16α (45 mg, 40%) as a powder. IR ν_{max} (KBr) 3289 (w), 3077 (w), 2924, 2853, 1627, 1556, 1467 cm^{-1} . 400 MHz $^{1}HNMR$ (CDCl₃-CD₃OD, 5:1) δ 0.86-0.90 (12H, m), 1.27 (66H, brs), 1.40-1.74 (12H, m), 1.97-2.02 (4H, m), 2.22-2.24 (2H, m), 3.22–4.13 (26H, m, containing two 3H, s, at δ 3.30 and 3.40 ppm), 4.77 (1H, d, J = 7.3 Hz), 5.35–5.39 (2H, m), 5.72 (1H, m). 125 MHz 13 C NMR (CDCl₃-CD₃OD, 1:1) δ 14.3 (4CH₃), 23.2 (4CH₂), 25.7 (CH₂), 26.3 (CH₂), 26.5 (CH₂), 26.8 (CH₂), 27.7 (2CH₂), 29.5 (CH₂), 29.9-30.4 (23CH₂), 31.0 (CH₂), 32.432.5 (4CH₂), 34.1 (CH₂), 34.4 (CH₂), 38.1 (CH₂), 55.7 (CH), 56.0 (CH₃), 59.3 (CH₃), 68.2 (CH₂), 69.1 (CH₂), 70.2 (CH₂ and CH), 71.3 (CH), 71.9 (CH₂), 74.0 (CH), 74.3 (CH₂), 74.9 (CH), 75.7 (CH), 79.1 (3CH), 81.6 (CH), 94.5 (CH), 101.5 (CH), 130.3 (CH), 130.4 (CH), 176.1 (CO). FABMS (negative-ion) m/z 1328 [M – H]⁻, 1350 [M + Na – 2H]⁻. HRFABMS Calcd for C₆₈H₁₃₂NO₁₉P₂: 1328.8871. Found: 1328.8873.

Phosphono 6-*O*-{2-deoxy-3-*O*-[(*R*)-3-(methoxy)decyl]-6-*O*-methyl-2-[(*Z*)-11-(octadecenoyl)amino]-4-*O*-phosphono- β -D-glucopyranosyl}-3-*O*-dodecyl-2-*O*-[(*R*)-3-(hydroxy)tetradecyl]- β -D-glucopyranoside (16 β). Compound 15 β (150 mg, 0.090 mmol) was treated as described for the formation of 16 α from 15 α to give 16 β (45 mg, 37%) as a powder. 400 MHz ¹H NMR (CDCl₃-CD₃OD, 5:1) δ 0.88 (12H, t, J = 6.6 Hz), 1.26 (66H, brs), 1.41–1.74 (12H, m), 2.00–2.04 (4H, m), 2.24–2.28 (2H, m), 3.12–4.07 (26H, m, containing two 3H, s, at δ 3.30 and 3.40 ppm), 4.92–4.97 (2H, m), 5.34–5.36 (2H, m). FABMS (negative-ion) m/z 1328 [M – H]⁻. Anal. Calcd for C₆₈H₁₃₃NO₁₉P₂: C, 61.38; H, 10.07; N, 1.05; P, 4.66%. Found: C, 61.22; H, 9.98; N, 1.10; P, 4.49%.

3-*O*-Decyl-1,2:5,6-di-*O*-isopropylidene-α-D-glucofuranose (17). To a solution of 1,2:5,6-di-O-isopropylidene- α -D-glucofuranose (17.00 g, 65.31 mmol) and decyl methanesulfonate (15.34 g, 64.90 mmol) in DMF (65 mL) was added NaH (55% oil dispersion, 3.40 g, 77.92 mmol, 1.5 equivalents) under ice-cooling temperature. After stirring for 15 min at 0 °C, the mixture was stirred for 16 h at room temperature. The reaction mixture was quenched with MeOH under ice-cooling temperature, and diluted with EtOAc. The solution was washed with water, and brine, dried over MgSO₄, filtered, and concentrated in vacuo to give a mixture, which was chromatographed on a silica-gel column. Elution with cyclohexane-EtOAc (6:1) gave 17 (22.50 g, 98%) as an oil. IR ν_{max} (film) 2987, 2928, 2857 cm⁻¹. 400 MHz ¹H NMR (CDCl₃) δ 0.88 (3H, t, J = 6.6 Hz), 1.27 (14H, brs), 1.32 (3H, s), 1.36 (3H, s), 1.43 (3H, s), 1.50 (3H, s), 1.55-1.58 (2H, m), 3.52 (1H, m), 3.59 (1H, m), 3.86 (1H, d, J = 3.7 Hz), 3.99 (1H, dd, J = 5.9, 8.8 Hz), 4.08 (1H, d, J = 5.9 Hz), 4.11 (1H, t, J = 6.6Hz), 4.14 (1H, m), 4.31 (1H, m), 4.53 (1H, d, J = 4.4 Hz), 5.88 (1H, d, J = 4.4 Hz). Anal. Calcd for $C_{22}H_{40}O_6$: C, 65.97; H, 10.07%. Found: C, 65.64; H, 9.94%.

3-O-decyl-4,6-O-isopropylidene- α , β -D-glucopyrano**side (18).** A solution of **17** (20.60 g, 51.43 mmol) obtained above in allyl alcohol (300 mL) containing 2% HCl was refluxed for 30 min, and concentrated in vacuo to give a mixture, which was dissolved in DMF (30 mL) and 2,2-dimethoxypropane (30 mL). To this solution was added p-TsOH•H₂O (500 mg). After stirring for 6 h at room temperature, the reaction mixture was diluted with EtOAc, which was washed with sat. aq NaHCO₃ and brine, dried over MgSO₄, filtered. The filtrate was concentrated in vacuo to give a mixture, which was chromatographed on a silica-gel column. Elution with cyclohexane-EtOAc (3:1) gave a 3:1 mixture of α - and β -anomer 18 (16.48 g, 80%) as an oil. The mixture was partly separated chromatographically. Physical data of α anomer: $R_f = 0.460$ (cyclohexane–EtOAc (3:1)); IR ν_{max} (film) 3470 (br), 2994, 2925, 2856 cm⁻¹. 400 MHz ¹H NMR (CDCl₃) δ 0.88 (3H, t, J = 6.6 Hz), 1.26 (14H, brs), 1.41 (3H, s), 1.49 (3H, s), 1.53–1.60 (2H, m), 2.27 (1H, d, J = 7.3 Hz, OH), 3.46-3.86 (8H, m), 4.04 (1H, m), 4.21 (1H, m), 4.93 (1H, d, J =3.7 Hz), 5.22-5.35 (2H, m), 5.93 (1H, m). Anal. Calcd for C₂₂H₄₀O₆: C, 65.97; H, 10.07%. Found: C, 65.71; H, 9.92%. Physical data of β -anomer: $R_f = 0.540$ (cyclohexane–EtOAc (3:1)); 400 MHz ¹H NMR (CDCl₃) δ 0.88 (3H, t, J = 6.6 Hz),

1.26 (14H, brs), 1.41 (3H, s), 1.49 (3H, s), 1.55–1.60 (2H, m), 2.29 (1H, d, J=2.2 Hz, OH), 3.25 (1H, m), 3.31 (1H, t, J=8.8 Hz), 3.45 (1H, m), 3.59–3.67 (2H, m), 3.75–3.82 (2H, m), 3.91 (1H, dd, J=5.1, 11.0 Hz), 4.14 (1H, dd, J=6.2, 12.8 Hz), 4.35 (1H, dd, J=5.1, 12.5 Hz), 4.40 (1H, d, J=8.1 Hz), 5.20–5.35 (2H, m), 5.93 (1H, m).

Allyl 3-O-decyl-4,6-O-isopropylidene-2-O-(3-oxotetradeca**noyl**)- α , β -D-glucopyranoside (19). To a solution of anomeric 3:1 mixture 18 (300 mg, 0.750 mmol) in CH₂Cl₂-THF (2:1, 6 mL) were added 3-oxotetradecanoic acid (242 mg, 1.00 mmol) and 1-[3-(dimethylamino)propyl]-3-ethylcarbodiimide hydrochloride (192 mg, 1.000 mmol). After stirring for 1 h at room temperature, the reaction mixture was diluted with CH2Cl2, washed with sat. aq NaHCO₃ and brine, dried over MgSO₄, and filtered. The filtrate was concentrated in vacuo to give a mixture, which was chromatographed on a silica-gel column. Elution with cyclohexane–EtOAc (6:1) gave a 3:1 mixture of α -anomer and β -anomer 19 (375 mg, 80%) as an oil. The mixture was partly separated chromatographically. Physical data of β -anomer: $R_f = 0.536$ (cyclohexane–EtOAc (4:1)); IR ν_{max} (film) 2923, 2853, 1752, 1722, 1636, 1568 cm⁻¹. 400 MHz ¹H NMR (CDCl₃) δ 0.88 (3H, t, J =6.6 Hz), 1.26 (30H, brs), 1.40 (3H, s), 1.41-1.68 (7H, m, containing 3H, s, at δ 1.49 ppm), 2.56 (2H, t, J = 6.3 Hz), 3.23 (1H, m), 3.38-3.56 (3H, m), 3.92 (1H, m), 4.05 (1H, m), 4.30 (1H, m), 4.47 (1H, d, J = 8.1 Hz), 4.95 (1H, m), 5.15-5.27 (2H, m),5.84 (1H, m). FABMS (positive-ion) m/z 623 $[M - H]^+$, 647 $[M + Na]^+$.

Allyl 3-O-decyl-2-O-(3-oxotetradecanovl)- α , β -D-glucopyra-A solution of anomeric 3:1 mixture **19** (2.37 g, 3.793 mmol) in MeOH (35 mL) containing p-toluenesulfonic acid monohydrate (200 mg) was stirred at room temperature for 2 h, and concentrated in vacuo to give a mixture, which was chromatographed on a silica-gel column. Elution with cyclohexane-EtOAc (3:2) gave a 3:1 mixture of α -anomer and β -anomer 20 (1.31 g, 59%) as an amorphous substance. The mixture was partly separated chromatographically. Physical data of α -anomer: $R_f = 0.346$ (cyclohexane–EtOAc (3:2)); 400 MHz 1 H NMR (CDCl₃) δ 0.88 (3H, t, J = 6.6 Hz), 1.26 (30H, brs), 1.51–1.60 (4H, m), 2.05 (2H, m, OH), 2.53 (2H, dt, J = 1.5, 6.6 Hz), 3.48 (2H, s), 3.59– 3.87 (7H, m), 4.01 (1H, m), 4.18 (1H, m), 4.74 (1H, dd, J = 3.7, 10.3 Hz), 5.08 (1H, d, J = 3.7 Hz, anomeric H), 5.19– 5.32 (2H, m), 5.89 (1H, m). Physical data of β -anomer: $R_f =$ 0.269 (cyclohexane–EtOAc (3:2)); IR ν_{max} (film) 3284 (br), 2921, 2852, 2596-2400 (w), 1752, 1720, 1649, 1632 cm⁻¹. 400 MHz ¹H NMR (CDCl₃) δ 0.88 (3H, t, J = 6.6 Hz), 1.26 (30H, brs), 1.51–1.63 (4H, m), 2.07 (1H, m, OH), 2.52–2.57 (2H, m), 3.20 (1H, d, J = 2.9 Hz), 3.45 (1H, s), 3.35–3.41 (2H, m), 3.58-3.68 (3H, m), 3.80 (1H, m), 3.91 (1H, m), 4.07 (1H, m), 4.31 (1H, m), 4.46 (1H, d, J = 8.1 Hz), 4.94 (1H, m), 5.16–5.28 (2H, m), 5.86 (1H, m). FABMS (positive-ion) m/z 583 $[M - H]^+$, 607 $[M + Na]^+$.

(E)-1-Propenyl 3-O-decyl-2-O-(3-oxotetradecanoyl)- α , β -D-glucopyranoside (21). To a solution of anomeric 3:1 mixture 20 (1.15 g, 1.966 mmol) in dry THF (40 mL) was added Ir[C₈H₁₂(MePh₂P)₂]PF₆ (25 mg). The atmospheric air in the reaction flask was replaced with nitrogen, then hydrogen to activate the iridium complex. After the red iridium complex turned colorless (ca. 30 sec) in solution, hydrogen was replaced with nitrogen. After stirring for 6 h under nitrogen at room temperature, the reaction mixture was concentrated in vacuo to give anomeric 3:1 mixture 21 (1.15 g, 100%), which was employed for the next reaction without purification. The mixture was partly separated

chromatographically. Elution with cyclohexane-EtOAc (3:2) gave the α -anomer ($R_f = 0.459$) as an amorphous compound and the β-anomer ($R_f = 0.324$) as a wax. Physical data of α-anomer: IR ν_{max} (KBr) 3408 (br), 2923, 2853, 1740, 1681 (w), 1659 (w) cm⁻¹. 400 MHz ¹H NMR (CDCl₃) δ 0.88 (3H, t, J = 6.6 Hz), 1.26 (30H, brs), 1.54–1.60 (7H, m, containing 3H, dd, J = 1.5, 6.6 Hz, at δ 1.55 ppm), 2.00 (1H, m, OH), 2.51–2.55 (2H, m), 3.48 (2H, s), 3.60-3.76 (5H, m), 3.80-3.83 (2H, m), 4.76 (1H, dd, J = 3.7, 9.5 Hz), 5.16 (1H, dd, J = 6.6, 12.5 Hz), 5.23 (1H, d, J = 3.7 Hz), 6.11 (1H, dd, J = 1.5, 12.5 Hz). Physical data of β -anomer: IR ν_{max} (film) 3500–3200 (br), 2922, 2853, 1745, 1720, 1682, 1648 (w), 1630 (w) cm⁻¹. 400 MHz ¹H NMR (CDCl₃) δ 0.88 (3H, t, J = 6.6 Hz), 1.26 (30H, brs), 1.51–1.62 (7H, m, containing 3H, d, J = 5.9 Hz at $\delta 1.54$ ppm), 2.02 (1H, brs, OH), 2.52–2.58 (2H, m), 3.19 (1H, d, J = 10.3 Hz), 3.36– 3.48 (3H, m, containing 2H, s, at δ 3.45 ppm), 3.57–3.70 (3H, m), 3.81 (1H, m), 3.93 (1H, m), 4.62 (1H, d, J = 8.1 Hz), 4.99 (1H, m), 5.10 (1H, m), 6.16 (1H, dd, J = 1.5, 12.5 Hz). FABMS (positive-ion) m/z 607 [M + Na]⁺.

(E)-1-Propenyl 6-O-tert-butyldimethylsilyl-3-O-decyl-2-O-(3-oxotetradecanoyl)- α , β -D-glucopyranoside (22). To a solution of anomeric 3:1 mixture 21 (2.89 g, 4.942 mmol) in CH₂Cl₂ (40 mL) were added tert-BuMe₂SiCl (800 mg, 5.315 mmol) and DMAP (650 mg, 5.315 mmol) for 16 h at room temperature. The reaction mixture was applied directly to a silica-gel column. Elution with cyclohexane-EtOAc (6:1) gave an anomeric 3:1 mixture 22 (3.45 g, 100%) as an oil. The mixture was partly separated chromatographically. Physical data of β -anomer: $R_f = 0.565$ (cylohexane/EtOAc = 4/1); IR ν_{max} (film) 3507 (br), 2927, 2856, 1754, 1721, 1682, 1663, 1621, 1465 cm⁻¹. 400 MHz ¹H NMR (CDCl₃) δ 0.08 (6H, m), 0.86–0.91 (15H, m, containing 9H, s, at δ 0.90 ppm), 1.25 (30H, brs), 1.50–1.61 (5H, m), 2.55– 2.58 (2H, m), 3.12-3.20 (1H, m), 3.36-3.46 (3H, m), 3.54-3.73 (3H, m), 3.83-3.93 (2H, m), 4.57 (1H, d, J = 8.8 Hz), 4.98(1H, m), 5.08 (1H, m), 6.14 (1H, d, J = 10.7 Hz). FABMS (positive-ion) m/z 697 [M – H]⁺, 721 [M + Na]⁺ (on addition of NaI).

(E)-1-Propenyl 4-O-allyloxycarbonyl-6-O-tert-butyldimethylsilyl-3-O-decyl-2-O-(3-oxotetradecanoyl)- α , β -D-glucopyranoside (23). To a solution of anomeric 3:1 mixture 22 (3.45 g, 4.935 mmol) in toluene (60 mL) were added pyridine (4.00 g, 50.569 mmol) and triphosgene (2.20 g, 7.414 mmol) 0 °C. After stirring for 15 min at 0 °C, allyl alcohol (6.90 g, 118.8 mmol) was added to this mixture. This solution was stirred for 1 h at 0 °C, and diluted with EtOAc. The solution was washed with sat. aq NaHCO₃ and brine, dried over MgSO₄, and filtered. The filtrate was concentrated in vacuo to give a mixture, which was chromatographed on a silica-gel column. Elution with cyclohexane-EtOAc (9:1) gave an anomeric 3:1 mixture 23 (2.63 g, 68%) as an oil. The mixture was partly separated chromatographically. Physical data of β -anomer: $R_f = 0.368$ (cylohexane/EtOAc = 9/1); IR ν_{max} (film) 2927, 2856, 1758, 1722, 1682 (w), 1664 (w), 1628 (w), 1464 cm⁻¹. 400 MHz ¹H NMR (CDCl₃) δ 0.04 (6H, s), 0.86-0.90 (15H, m, containing 9H, s, at 0.88 ppm), 1.26 (30H, brs), 1.42–1.60 (7H, m, containing 3H, d, J = 7.3Hz, at 1.52 ppm), 2.54 (2H, t, J = 7.3 Hz), 3.19 (1H, d, J = 8.8Hz), 3.43 (1H, s), 3.47-3.57 (4H, m), 3.72-3.73 (2H, m), 4.57 (1H, d, J = 8.1 Hz), 4.61–4.65 (2H, m), 4.78 (1H, t, J = 9.5Hz), 4.97-5.14 (2H, m), 5.27-5.39 (2H, m), 5.92 (1H, m), 6.16 (1H, dd, J = 1.5, 12.5 Hz). FABMS (positive-ion) m/z 805 $[M + Na]^+$.

4-*O*-Allyloxycarbonyl-3-*O*-decyl-2-*O*-(3-oxotetradecanoyl)- α , β -D-glucopyranose (24). To a solution of anomeric 3:1 mix-

ture **23** (2.20 g, 4.935 mmol) in CH₂Cl₂ (30 mL) and MeCN (60 mL) was added an aq 48% HF solution (20 mL). The mixture was stirred vigorously for 1.5 days at room temperature, and diluted with EtOAc, which was washed with H₂O, sat. aq NaHCO₃ and brine, dried over MgSO₄, and filtered. The filtrate was concentrated in vacuo to give a residue, which was chromatographed on a silica-gel column. Elution with cyclohexane–EtOAc (2:3) gave an anomeric mixture **24** (1.02 g, 58%) as an oil. IR ν_{max} (film) 3430, 2925, 2855, 1754, 1719, 1650 (w), 1466 cm⁻¹. 400 MHz ¹HNMR (CDCl₃) δ 0.88 (6H, t, J = 6.6 Hz), 1.26 (30H, brs), 1.45–1.70 (4H, m), 2.50–2.57 (2H, m), 3.47–3.80 (7H, m), 3.90 (1H, t, J = 9.5 Hz), 4.02 (1H, m), 4.66 (2H, d, J = 5.9 Hz), 4.76–4.88 (1.5H, m), 5.29–5.40 (2H, m), 5.49 (0.5H, brs), 5.93 (1H, m). FABMS (positive-ion) m/z 369, 611, 651 [M + Na]⁺. HRFABMS Calcd for $C_{34}H_{60}O_{10}Na$: 651.4085. Found: 651.4052.

4-O-Allyloxycarbonyl-3-O-decyl-6-O-{2-deoxy-4-O-diallylphosphono-3-O-[(R)-3-(methoxy)decvl]-6-O-methyl-2-[(trichloroethoxycarbonyl)amino]- β -D-glucopyranosyl}-2-O-(3-oxotetradecanoyl)-D-glucopyranose (25). Compound **24** (641 mg, 1.019 mmol) and imidate 11 (877 mg, 1.040 mmol) were treated as described for the formation of 12 from 10 and 11 to give 25 (1.300 g, 97%) as a gum. IR ν_{max} (film) 3500–3200, 2927, 2856, 1753, 1737, 1721 (shoulder) cm⁻¹. 400 MHz ¹H NMR (CDCl₃) 0.88 (9H, t, J = 6.6 Hz), 1.26 (40H, brs), 1.40–1.50 (4H, m), 1.57–1.63 (2H, m), 1.70–1.78 (2H, m), 2.51–2.55 (2H, m), 3.35–3.88 (24H, m, containing two 3H, s, at δ 3.27 and 3.39 ppm), 4.18-4.30 (2H, m), 4.55 (0.5H, m), 5.25-5.43 (6.5H, m), 5.89-5.99 (3H, m), 6.65 (1H, br). FABMS (positive-ion) m/z 1330 [M + Na, 35 Cl]⁺, 1332. HRFABMS Calcd for C₆₁H₁₀₅³⁵Cl₃NO₂₀PNa: 1330.5931. Found: 1330.5928.

4-O-Allyloxycarbonyl-3-O-decyl-6-O-{2-deoxy-4-O-diallylphosphono-3-O-[(R)-3-(methoxy)decyl]-6-O-methyl-2-[(Z)-11-(octadecenoyl)amino]- β -D-glucopyranosyl}-2-O-(3-oxotetradecanoyl)-D-glucopyranose (26). To a solution of 25 (1.290 g, 0.985 mmol) in THF-AcOH (1:1, 20 mL) was added Zn dust (2.00 g, 30.595 m atom). The mixture was stirred vigorously with a magnetic stirrer at 25 °C for 3 h, and filtered. The filtrate was concentrated in vacuo below 30 °C, and diluted with EtOAc, which was washed with sat. aq NaHCO3, and brine, dried over MgSO₄, and filtered. The filtrate was concentrated in vacuo to give amine, which was dissolved in THF (6 mL)-H₂O (2 mL) containing NaHCO₃ (250 mg, 2.976 mmol). To this solution was added a solution of cis-vaccenyl chloride (356 mg, 1.186 mmol, 1.2 equivalents) in THF (2 mL) with vigorous stirring at 25 °C. (This acid chloride was prepared from (Z)-11-octadecenoic acid and excess oxalyl chloride in benzene at room temperature for 3 h). After stirring for 2 h, the reaction mixture was diluted with EtOAc, which was washed with sat. aq NaHCO3 and brine, dried over MgSO₄, and filtered. The filtrate was concentrated in vacuo to give a mixture, which was chromatographed on a silica-gel column. Elution with EtOAc-hexane (3:1) gave 26 (756 mg, 55%) as an oil. IR ν_{max} (film) 3305 (br), 2926, 2856, 1755, 1652, 1543, 1465 cm⁻¹. 400 MHz ¹HNMR (CDCl₃) δ 0.88 (12H, m), 1.26 (60H, brs), 1.40-1.80 (10H, m), 2.01-2.02 (4H, m), 2.16-2.22 (2H, m), 2.52-2.59 (2H, m), 3.04-4.37 (25H, m, containing two 3H, s, at δ 3.27 and 3.39 ppm), 4.54– 4.78 (7H, m), 5.00 (0.5H, s), 5.23-5.52 (8.5H, m), 5.86-5.98 (3H, m), 6.09 (1H, d, J = 7.3 Hz, NH). FABMS (positive-ion) m/z 1420 [M + Na]⁺. HRFABMS Calcd for C₇₆H₁₃₆NO₁₉PNa: 1420.9342. Found: 1420.9347.

Diallylphosphono 4-*O*-allyloxycarbonyl-3-*O*-decyl-6-*O*-{2-deoxy-4-*O*-diallylphosphono-3-*O*-[(*R*)-3-(methoxy)decyl]-6-*O*-

methyl-2-[(Z)-11-(octadecenoyl)amino]- β -D-glucopyranosyl}-2-O-(3-oxotetradecanoyl)- α -D-glucopyranoside (27 α) and Diallylphosphono 4-O-allyloxycarbonyl-3-O-decyl-6-O-{2-deoxy-4-*O*-diallylphosphono-3-*O*-[(*R*)-3-(methoxy)decyl]-6-*O*-methyl-2-[(Z)-11-(octadecenoyl)amino]- β -D-glucopyranosyl}-2-O-(3oxotetradecanoyl)- β -D-glucopyranoside (27 β). The aboveobtained anomeric mixture 26 (743 mg, 0.531 mmol) was treated as described for the formation of 15β and 15α from 14 to give 27β (166 mg, 20%) [$R_f = 0.448$, cyclohexane–EtOAc (1:2)] and 27α (185 mg, 22%) [$R_f = 0.299$, cyclohexane–EtOAc (1:2)] as an oil. Physical data of 27β : IR ν_{max} (film) 3309, 3086 (w), 2926, 2856, 1759, 1722, 1669, 1550 cm $^{-1}$. 400 MHz 1 H NMR (CDCl₃) δ 0.88 (12H, t, J = 6.6 Hz), 1.26 (60H, brs), 1.38-1.70 (10H, m), 2.00-2.04 (4H, m), 2.21-2.23 (2H, m), 2.53 (2H, t, J = 7.4 Hz), 3.21-3.79 (21H, m, containing two 3H, s, at δ 3.27 and 3.38 ppm), 3.98 (1H, m), 4.27 (1H, m), 4.46–4.72 (12H, m), 5.00–5.09 (2H, m), 5.23-5.42 (12H, m), 5.85-5.98 (5H, m), 7.35 (1H, d, J = 9.5Hz, NH). FABMS (positive-ion) m/z 1580 [M + Na]⁺. HRFABMS Calcd for $C_{82}H_{145}NO_{22}P_2Na$: 1580.9632. Found: 1580.9650. Physical data of 27α : IR ν_{max} (film) 3306, 3086 (w), 2926, 2856, 1759, 1721, 1666, 1548 cm⁻¹. 400 MHz ¹H NMR (CDCl₃) δ 0.88 (12H, t, J = 6.6 Hz), 1.26 (60H, brs), 1.46–1.73 (10H, m), 2.00-2.05 (4H, m), 2.21-2.24 (2H, m), 2.53 (2H, t, J =7.7 Hz), 3.24–3.79 (19H, m, containing two 3H, s, at δ 3.28 and 3.38 ppm), 3.88–3.98 (2H, m), 4.08 (1H, m), 4.27 (1H, m), 4.54-4.65 (11H, m), 4.75-4.83 (2H, m), 5.24-5.42 (12H, m), 5.80 (1H, dd, J = 3.7, 6.6 Hz), 5.89–5.99 (5H, m), 6.54 (1H, d, J = 7.3 Hz, NH). FABMS (positive-ion) m/z 1558 [M + H]⁺, 1580 $[M + Na]^+$. HRFABMS Calcd for $C_{82}H_{145}NO_{22}P_2Na$: 1580.9632. Found: 1580.9617.

Phosphono 3-O-decyl-6-O- $\{2$ -deoxy-3-O- $\{(R)$ -3-(methoxy)decyl]-6-O-methyl-2-[(Z)-11-(octadecenoyl)amino]-4-O-phosphono- β -D-glucopyranosyl}-2-O-(3-oxotetradecanoyl)- α -D-glucopyranoside (28 α). Compound 27 α (178 mg, 0.114 mmol) was treated as described for the formation of 16α from 15α to give **28α** (89 mg, 59%) as a powder. IR ν_{max} (KBr) 3286 (br), 2925, 2854, 1747, 1717, 1630, 1551, 1466 cm⁻¹. 400 MHz ¹H NMR (CDCl₃-CD₃OD, 5:1) δ 0.88 (12H, t, J = 6.6 Hz), 1.27 (60H, brs), 1.40-1.75 (10H, m), 2.00-2.04 (4H, m), 2.22-2.25 (2H, m), 2.58 (2H, t, J = 7.3 Hz), 3.24-4.10 (24H, m), 4.69 (1H, m), 4.76 (1H, d, J = 8.1 Hz), 5.33-5.36 (2H, m), 5.67 (1H, m). 125MHz 13 C NMR (CDCl₃-CD₃OD, 1:1) δ 14.3 (4CH₃), 23.2 (4CH₂), 24.0 (CH₂), 25.7 (CH₂), 26.5 (CH₂), 26.6 (CH₂), 27.7 (2CH₂), 29.5-30.4 (21CH₂), 30.9 (CH₂), 32.3 (CH₂), 32.4 (CH₂), 32.5 (2CH₂), 34.1 (CH₂), 34.4 (CH₂), 37.1 (CH₂), 43.5 (CH₂), 55.8 (CH), 56.0 (CH₃), 59.3 (CH₃), 68.1 (CH₂), 70.2 (CH and CH₂), 71.8 (CH₂), 73.9 (CH), 74.1 (CH₂), 74.8 (CH), 75.7 (CH), 79.1 (3CH), 81.4 (CH), 93.5 (CH), 101.5 (CH), 130.3 (CH), 130.4 (CH), 167.7 (CO), 176.2 (CO), 204.1 (CO). FABMS (negative-ion) m/z 1312 [M – H]⁻. Anal. Calcd for C₆₆H₁₂₅NO₂₀P₂: C, 60.30; H, 9.58; N, 1.07; P, 4.71%. Found: C, 60.20; H, 9.49; N, 1.20; P, 4.69%.

Phosphono 3-*O*-decyl-6-*O*-{2-deoxy-3-*O*-[(*R*)-3-(methoxy)-decyl]-6-*O*-methyl-2-[(*Z*)-11-(octadecenoyl)amino]-4-*O*-phosphono-β-D-glucopyranosyl}-2-*O*-(3-oxotetradecanoyl)-β-D-glucopyranoside (28β). Compound 27β (160 mg, 0.103 mmol) was treated as described for the formation of 16α from 15α to give 28β (97 mg, 72%) as a powder. IR ν_{max} (KBr) 3287 (br), 2925, 2854, 1746, 1717, 1629, 1554, 1466 cm⁻¹. 400 MHz ¹H NMR (CDCl₃-CD₃OD, 5:1) δ 0.88 (12H, t, J = 6.6 Hz), 1.26 (60H, brs), 1.40–1.73 (10H, m), 2.01–2.02 (4H, m), 2.23–2.27 (2H, m), 2.58 (2H, t, J = 7.3 Hz), 3.30–3.82 (21H, m, containing two 3H,

s, at 3.30 and 3.41 ppm), 4.01–4.05 (3H, m), 4.88–4.94 (2H, m), 5.05 (1H, m), 5.33–5.36 (2H, m). 125 MHz $^{13}\mathrm{C}$ NMR (CDCl₃–CD₃OD, 1:1) δ 14.3 (4CH₃), 23.1 (4CH₂), 23.9 (CH₂), 25.6 (CH₂), 26.5 (CH₂), 27.6 (CH₂), 27.7 (2CH₂), 29.4–30.3 (21CH₂), 30.7 (CH₂), 32.3 (2CH₂), 32.4 (2CH₂), 34.0 (CH₂), 34.3 (CH₂), 36.9 (CH₂), 43.5 (CH₂), 55.2 (CH), 55.9 (CH₃), 59.2 (CH₃), 67.7 (CH₂), 70.1 (CH₂), 70.6 (CH), 71.8 (CH₂), 73.8 (CH₂), 74.5 (CH), 75.6 (CH), 79.0 (CH), 79.2 (3CH), 82.7 (CH), 96.6 (CH), 101.6 (CH), 130.2 (CH), 130.3 (CH), 166.8 (CO), 175.9 (CO), 204.0 (CO). FABMS (negative-ion) m/z 1312 [M – H]⁻. Anal. Calcd for C₆₆H₁₂₅NO₂₀P₂: C, 60.30; H, 9.58; N, 1.07; P, 4.71%. Found: C, 60.18; H, 9.74; N, 1.11; P, 4.58%.

Allyl 4,6-*O*-isopropylidene-α-D-glucopyranoside (30). A solution of **29** (5.00 g, 22.7 mmol) in DMF (10 mL) and 2,2-dimethoxypropane (15 mL) containing *p*-TsOH·H₂O (200 mg) was stirred for 16 h at room temperature. The reaction mixture was diluted with EtOAc, which was washed with a small amount of sat. aq NaHCO₃-brine, dried over MgSO₄, and filtered. The filtrate was concentrated in vacuo to give **30** (3.84 g, 65%) as an oil. IR ν_{max} (film) 3434 (br) cm⁻¹. 400 MHz ¹H NMR (CDCl₃) δ 1.45 (3H, s), 1.52 (3H, s), 2.30 (1H, d, J = 10.3 Hz, OH), 2.79 (1H, d, J = 2.2 Hz, OH), 3.51–3.88 (6H, m), 4.03 (1H, m), 4.23 (1H, m), 4.92 (1H, d, J = 3.7 Hz), 5.23–5.34 (2H, m), 5.91 (1H, m).

Allyl 2,3-di-*O*-dodecyl-4,6-*O*-isopropylidene-α-D-glucopyranoside (31). To a solution of 30 (2.90 g, 11.14 mmol) and dodecyl iodide (8.00 g, 27.00 mmol) in DMF (29 mL) was added NaH (55% oil dispersion, 2.36 g, 54.00 mmol) at room temperature, and the mixture was stirred for 16 h at room temperature. The reaction mixture was poured into ice water, and extracted with EtOAc, which was washed with brine, dried over MgSO₄, and filtered. The filtrate was concentrated to give a mixture, which was chromatographed on a silica-gel column. Elution with cyclohexane–EtOAc (9:1) gave 31 (6.49 g, 98%) as an oil. 400 MHz 1 H NMR (CDCl₃) δ 0.88 (6H, t, J = 6.6 Hz), 1.26 (36H, brs), 1.41 (3H, s), 1.48 (3H, s), 1.50–1.60 (4H, m), 3.30 (1H, m), 3.49–3.74 (8H, m), 3.83 (1H, m), 4.08 (1H, dd, J = 6.6, 13.3 Hz), 4.18 (1H, dd, J = 5.1, 13.3 Hz), 4.90 (1H, d, J = 3.7 Hz), 5.22 (1H, m), 5.32 (1H, m), 5.92 (1H, m).

Allyl 2,3-di-O-dodecyl- α -D-glucopyranoside (32). To a solution of 31 (6.40 g, 10.72 mmol) in MeOH (50 mL) was added TsOH·H₂O (300 mg, 1.58 mmol). The mixture was stirred for 40 min at room temperature, and diluted with EtOAc, which was washed with sat. aq NaHCO₃ and brine, dried over MgSO₄, and filtered. The filtrate was concentrated in vacuo to give a residue, which was chromatographed on a silica-gel column. Elution with cyclohexane-EtOAc (1:1) gave 32 (3.23 g, 54%) as a solid. IR ν_{max} (KBr) 3364 (br), 2919, 2850, 1467 cm⁻¹. 400 MHz ¹H NMR (CDCl₃) δ 0.88 (6H, t, J = 6.6 Hz), 1.26 (36H, brs), 1.54-1.60 (4H, m), 1.95 (1H, t, J = 5.7 Hz, OH), 2.46 (1H, d, J =2.9 Hz, OH), 3.28 (1H, m), 3.46-3.90 (8H, m), 3.93 (1H, m), 4.07 (1H, dd, J = 6.6, 13.2 Hz), 4.20 (1H, dd, J = 5.1, 13.2 Hz), 4.96(1H, d, J = 3.7 Hz), 5.21-5.35 (2H, m), 5.93 (1H, m). FABMS (positive-ion) m/z 555 [M – H]⁺, 579 [M + Na]⁺. HRFABMS Calcd for $C_{33}H_{64}O_6Na$: 579.4600. Found: 579.4581.

(E)-1-Propenyl 2,3-di-O-dodecyl-α-D-glucopyranoside (33). To a solution of 32 (2.602 g, 4.672 mmol) in dry THF (40 mL) was added (1,5-cyclooctadiene)bis(methyldiphenylphosphine)-iridium(I) hexafluorophosphate (50 mg). The atmosphere was replaced with nitrogen, and then hydrogen to activate the iridium complex. After the red iridium complex turned colorless in solu-

tion by stirring for about 10 sec under a hydrogen atmosphere, the hydrogen was replaced again by nitrogen. The solution was stirred for 16 h at room temperature, and concentrated in vacuo to give **33** (quantitatively) as a solid. IR $\nu_{\rm max}$ (KBr) 3341 (br), 2919, 2851, 1678 (w), 1468 cm⁻¹. 400 MHz ¹H NMR (CDCl₃) δ 0.88 (6H, t, J = 6.6 Hz), 1.26 (36H, brs), 1.55–1.60 (9H, m), 1.89 (1H, brs, OH), 2.50 (1H, brs, OH), 3.30 (1H, dd, J = 3.7, 8.8 Hz), 3.46–3.78 (6H, m), 3.70–3.84 (2H, m), 3.93 (1H, m), 5.12 (1H, d, J = 3.7 Hz), 5.19 (1H, qd, J = 6.6, 12.5 Hz), 6.18 (1H, dd, J = 1.5, 12.5 Hz). FABMS (positive-ion) m/z 555 [M – H]⁺, 579 [M + Na]⁺.

(E)-1-Propenyl 6-*O-tert*-butyldimethylsilyl-2,3-di-*O*-dodecyl- α -D-glucopyranoside (34). To a solution of 33 (2.700 g, 4.848 mmol) in CH₂Cl₂ (50 mL) were added tert-BuMe₂SiCl (880 mg, 5.838 mmol) and DMAP (713 mg, 5.838 mmol). The solution was stirred for 16 h at room temperature. The mixture was directly chromatographed on a silica-gel column. Elution with cyclohexane-EtOAc (7:1, and then 2:1) gave 34 (3.105 g, 95%) as an oil. IR ν_{max} (film) 3504 (br), 2926, 2855, 1680 (w), 1659 (w), 1464 cm^{-1} . $400 \text{ MHz} ^{1}\text{H NMR}$ (CDCl₃) δ 0.08 (6H, s), 0.88 (15H, m, containing 9H, s, at δ 0.90 ppm), 1.26 (36H, brs), 1.58-1.65 (7H, m), 2.71 (1H, d, J = 2.2 Hz, OH), 3.28 (1H, dd, J = 2.7, 9.5 Hz), 3.52–3.90 (9H, m), 5.10 (1H, d, J = 3.7 Hz), 5.19 (1H, m), 6.19 (1H, dd, J = 2.2, 12.5 Hz). FABMS (positive-ion) m/z 671 [M + H]⁺, 693 [M + Na]⁺ (on addition of NaI). HRFABMS Calcd for C₃₉H₇₈O₆Si: 693.5466. Found: 693.5469.

(E)-1-Propenyl 4-O-allyloxycarbonyl-6-O-tert-butyldimethylsilyl-2,3-di-O-dodecyl- α -D-glucopyranoside (35). To a solution of 34 (3.10 g, 4.619 mmol) in toluene (45 mL) and pyridine (3.5 mL, 3.423 g) was added triphosgene (1.60 g, 5.392 mmol) at 0 °C. After stirring for 10 min, allyl alcohol (6.00 g) was added to this solution. The mixture was stirred for 1 h at 0 °C, and diluted with EtOAc, which was washed with sat. aq NaHCO₃ and brine, dried over MgSO₄, and filtered. The filtrate was concentrated in vacuo to give a residue, which was chromatographed on a silica-gel column. Elution with cyclohexane-EtOAc (14:1) gave **35** (3.49 g, 100%) as an oil. IR ν_{max} (film) 2926, 2856, 1758, 1680 (w), 1661 (w), 1464 cm⁻¹. 400 MHz ¹H NMR (CDCl₃) δ 0.02 (3H, s), 0.03 (3H, s), 0.88 (9H, s), 0.88 (6H, t, J = 6.6Hz), 1.25 (36H, brs), 1.51–1.57 (7H, m), 3.35 (1H, dd, J = 3.7, 9.7 Hz), 3.51–3.83 (8H, m), 4.62–4.65 (2H, m), 4.72 (1H, t, J =9.5 Hz), 5.19 (1H, d, J = 3.7 Hz), 6.19 (1H, m), 5.26–5.39 (2H, m), 5.94 (1H, m), 6.18 (1H, dd, J = 1.5, 12.5 Hz). FABMS (positive-ion) m/z 753 [M – H]⁺, 777 [M + Na]⁺ (on addition of NaI). HRFABMS Calcd for $C_{43}H_{82}O_8SiNa$: 777.5677. Found: 777.5641.

4-*O*-**Allyloxycarbonyl-2,3-di-***O*-**dodecyl-D**-**glucopyranose** (**36**). To a solution of **35** (3.60 g, 4.767 mmol) in CH₂Cl₂ (10 mL) and MeCN (20 mL) was added aq 48% HF solution (3 mL). The mixture was stirred for 16 h at room temperature, and diluted with EtOAc, which was washed with H₂O, sat. aq NaHCO₃ and brine, dried over MgSO₄, and filtered. The filtrate was concentrated in vacuo to give a residue, which was chromatographed on a silica-gel column. Elution with EtOAc gave an anomeric mixture **36** (2.12 g, 74%) as an oil. IR ν_{max} (film) 3446 (br), 2920, 2851, 1754, 1724, 1468 cm⁻¹. 400 MHz ¹H NMR (CDCl₃) δ 0.88 (6H, t, J = 6.6 Hz), 1.26 (32H, brs), 1.43–1.58 (2H, m), 1.69 (1H, brs, OH), 3.12–3.98 (7H, m), 4.65–4.72 (3H, m), 5.28–5.40 (2H, m), 5.93 (1H, m). FABMS (positive-ion) m/z 583 [M + Na]⁺. HRFABMS Calcd for C₃₄H₆₄O₈Na: 623.4499. Found: 623.4490.

4-*O*-Allyloxycarbonyl-2,3-di-*O*-dodecyl-6-*O*-{2-deoxy-4-*O*-diallylphosphono-3-*O*-[(*R*)-3-(methoxy)decyl]-6-*O*-methyl-2-[(2,2,2-trichloroethoxycarbonyl)amino]- β -D-glucopyranosyl}-D-glucopyranose (37). Compound 36 (1.82 g, 3.112 mmol) and imidate 11 (2.65 g, 3.142 mmol) were treated as described for the formation of 12 from 10 and 11 to give an anomeric mixture 37 (3.86 g, 96%) as a gum. IR ν_{max} (film) 3500–3100, 2926, 2856, 1753, 1736, 1542, 1465 cm⁻¹. 400 MHz ¹H NMR (CDCl₃) 0.88 (9H, t, *J* = 6.6 Hz), 1.26 (48H, brs), 1.43–1.75 (6H, m), 3.11–3.90 (24H, m), 4.13–4.40 (2H, m), 4.55–4.85 (10H, m), 5.24–5.39 (6H, m), 5.89–5.97 (3H, m). FABMS (positive-ion) *m/z* 1304, 1302 [M + Na, ³⁵Cl]⁺, 1332. HRFABMS Calcd for C₆₁H₁₀₉³⁵Cl₃NO₁₈Na: 1302.6346. Found: 1302.6333.

4-*O*-Allyloxycarbonyl-6-*O*-{2-deoxy-4-*O*-diallylphosphono-3-*O*-[(*R*)-3-(methoxy)decyl]-6-*O*-methyl-2-[(*Z*)-11-(octadecenoyl)amino]-β-D-glucopyranosyl}-2,3-di-*O*-dodecyl-D-glucopyranose (38). Compound 37 (1.90 g, 1.482 mmol) was treated as described for the formation of 26 from 25 to give 38 (1.32 g, 65%) as an oil. IR $\nu_{\rm max}$ (film) 3302 (br), 2926, 2855, 1754, 1652, 1549, 1465 cm⁻¹. 400 MHz ¹H NMR (CDCl₃) δ 0.88 (12H, m), 1.26 (66H, brs), 1.40–1.80 (10H, m), 1.99–2.02 (4H, m), 2.17–2.33 (2H, m), 3.10–4.10 (25H, m), 4.24–4.63 (8H, m), 5.21–5.32 (8H, m), 5.87–5.98 (3H, m), 6.12 (1H, m, NH). FABMS (positive-ion) m/z 1392 [M + Na]⁺. HRFABMS Calcd for C₇₆H₁₄₀NO₁₇PNa: 1392.9756. Found: 1392.9774.

Diallylphosphono 4-O-allyloxycarbonyl-6-O-{2-deoxy-4-Odiallylphosphono-3-O-[(R)-3-(methoxy)decyl]-6-O-methyl-2-[(Z)-11-(octadecenovl)amino]- β -D-glucopyranosyl}-2,3-di-Ododecyl-\alpha-D-glucopyranoside (39\alpha) and Diallylphosphono 4-O-allyloxycarbonyl-6-O-{2-deoxy-4-O-diallylphosphono-3-O-[(R)-3-(methoxy)decyl]-6-O-methyl-2-[(Z)-11-(octadecenoyl)amino]- β -D-glucopyranosyl}-2,3-di-O-dodecyl- β -D-glucopyra**noside** (39 β). The above-obtained anomeric mixture 38 (1.32 g, 0.963 mmol) was treated as described for the formation of 15β and 15 α from 14 to give 39 β (378 mg, 26%, $R_f = 0.380$ [cyclohexane-EtOAc (1:1)]) as an oil and 39 α (356 mg, 25%, $R_f =$ 0.240 (cyclohexane-EtOAc (1:1)) as an oil. Physical data of **39** β : IR ν_{max} (film) 3310 (w), 3086 (w), 2926, 2855, 1757, 1668, 1549, 1465 cm⁻¹. 400 MHz ¹H NMR (CDCl₃) δ 0.88 (12H, t, J = 6.6 Hz), 1.26 (66H, brs), 1.40–1.75 (10H, m), 2.00-2.02 (4H, m), 2.10-2.25 (2H, m), 3.18-3.75 (22H, m, containing two 3H, s, at δ 3.27 and 3.38 ppm), 4.00 (1H, m), 4.25 (1H, m), 4.47-4.63 (11H, m), 4.70 (1H, d, J = 8.8 Hz), 4.95(1H, dd, J = 5.1, 8.1 Hz), 5.23–5.43 (12H, m), 5.86–5.97 (5H, m), 7.48 (1H, d, J = 9.5 Hz, NH). FABMS (positive-ion) m/z1552 $[M + Na]^+$. HRFABMS Calcd for $C_{82}H_{149}NO_{20}P_2Na$: 1553.0046. Found: 1553.0040. Physical data of 39α : IR ν_{max} (film) 3310, 3085, 2926, 2855, 1757, 1665, 1548, 1465 cm⁻¹. 400 MHz ¹H NMR (CDCl₃) δ 0.88 (12H, t, J = 6.6 Hz), 1.26 (66H, brs), 1.40-1.75 (10H, m), 1.99-2.01 (4H, m), 2.21-2.25 (2H, m), 3.21–3.83 (22H, m, containing two 3H, s, at δ 3.28 and 3.38 ppm), 3.96–4.02 (2H, m), 4.26 (1H, dd, J = 9.5, 18.3 Hz), 4.52–4.73 (11H, m), 5.23–5.42 (12H, m), 5.73 (1H, dd, J = 3.3, 6.3 Hz), 5.87–6.01 (5H, m), 6.53 (1H, d, J = 8.1 Hz, NH). FABMS (positive-ion) m/z 1552 [M + Na]⁺. HRFABMS Calcd for C₈₂H₁₄₉NO₂₀P₂Na: 1553.0046. Found: 1553.0041.

Phosphono 6-*O*-{2-deoxy-3-*O*-[(*R*)-3-(methoxy)decyl]-6-*O*-methy-4-*O*-phosphono-2-[(*Z*)-11-(octadecenoyl)amino]- β -p-glucopyranosyl}-2,3-di-*O*-dodecyl- α -p-glucopyranoside (40 α). Compound 39 α (153 mg, 0.100 mmol) was treated as described for the formation of 16 α from 15 α to give 40 α (70 mg, 54%) as a powder. IR ν_{max} (KBr) 3292 (br), 2924, 2853, 1630, 1551,

1467, 1377 cm⁻¹. 400 MHz ¹H NMR (CDCl₃–CD₃OD, 5:1) δ 0.88 (12H, t, J=6.6 Hz), 1.27 (66H, brs), 1.40–1.76 (10H, m), 1.96–2.04 (4H, m), 2.22–2.26 (2H, m), 3.23–4.10 (25H, m, containing two 3H, s, at δ 3.30 and 3.40 ppm), 4.74 (1H, d, J=8.3 Hz), 5.34–5.39 (2H, m), 5.64 (1H, m). FABMS (negative-ion) m/z 1284 [M – H]⁻. HRFABMS Calcd for C₆₆H₁₂₈NO₁₈P₂: 1284.8607. Found: 1284.8601. Anal. Calcd for C₆₆H₁₂₉NO₁₈P₂: C, 61.61; H, 10.11; N, 1.09; P, 4.81%. Found: C, 61.77; H, 9.96; N, 1.10; P, 4.73%.

Phosphono 6-*O*-{2-deoxy-3-*O*-[(*R*)-3-(methoxy)decyl]-6-*O*-methy-4-*O*-phosphono-2-[(*Z*)-11-(octadecenoyl)amino]- β -D-glucopyranosyl}-2,3-di-*O*-dodecyl- β -D-glucopyranoside (40 β). Compound 39 β (185 mg, 0.1210 mmol) was treated as described for the formation of 16α from 15α to give 40 β (106 mg, 68%) as a powder. IR ν_{max} (KBr) 3387, 3299, 2924, 2853, 1631, 1552, 1467, 1377 cm⁻¹. 400 MHz ¹H NMR (CDCl₃-CD₃OD, 4:1) δ 0.89 (12H, t, *J* = 6.6 Hz), 1.27 (66H, brs), 1.40–1.76 (10H, m), 2.00–2.04 (4H, m), 2.25–2.29 (2H, m), 3.11–3.89 (23H, m, containing two 3H, s, at δ 3.31 and 3.41 ppm), 4.00–4.05 (2H, m), 4.94–4.98 (2H, m), 5.32–5.37 (2H, m). FABMS (negative-ion) m/z 1284 [M – H]⁻. HRFABMS Calcd for C₆₆H₁₂₈NO₁₈P₂: 1284.8607. Found: 1284.8601.

Methods for Measurement of Biological Activity. The sources of the materials used in the study were as follows: lipopolysaccharide (LPS) from $E.\ coli$ serotype 026:B6 and 12-O-tetradecanoylphorbol acetate (TPA) were from Sigma, St. Louis, MO; RPMI-1640 medium, fetal bovine serum (FBS), and newborn calf serum (NBCS) were from Gibco, Grand Island, NY; and human TNF α ELISA kit and mouse TNF α ELISA kit were from Genzyme-Techne, Minneapolis, MN.

Production of TNF α by Human Whole Blood.⁹ Materials: Lipopolysaccharide (LPS, lot 50K4117, *E. coli* 026:B6), human tumor necrosis factor alpha (TNF α) immunoassay kit and 96-well assay plates were purchased from Sigma, BioSource International, Inc. and Corning Inc. (Cat. No. 3956), respectively.

Whole Blood TNF α Production: Fresh blood was collected aseptically in the presence of heparin by venipuncture from healthy adult volunteers. The subjects did not have any apparent inflammatory conditions and had taken no drugs for at least 7 days prior to blood collection. Written informed consent was obtained from all volunteers before the experiment. In each well of the plates, 360 µL aliquots of blood were mixed with 20 µL of LPS solution (200 ng/mL) dissolved in PBS in the presence (for test sample) or absence (for positive control sample) of test compounds solution (dissolved in 10% DMSO/PBS solution). For the negative control samples, the same amount of blood was cultured without either LPS or a test compound solution. After 6 h of incubation at 37 °C, the plates were centrifuged at $490 \times g$ for 15 min, and the plasma was collected and stored at -20 °C. The concentrations of TNF α in the plasma were measured with commercially available immunoassay kits.

Statistical Analysis: The percentage of inhibition of TNF α production was calculated by the following formula: $[1-(concentration of TNF\alpha in the test sample - concentration of TNF<math>\alpha$ in the negative control sample)/(concentration of TNF α in the positive control sample - concentration of TNF α in the negative control sample)] × 100. The suppressive activities of test compounds are expressed as the fifty percent inhibitory concentration (IC₅₀) of the test compound, the concentration at which the test compound suppresses TNF α production by 50%. The IC₅₀ was calculated from the percentage of inhibition using the SAS System for Windows (version 5). The results are expressed as the mean

IC₅₀ of triplicate experiments.

Production of TNFα by Galactosamine Loaded C3H/HeN Mice. ¹⁰ Materials; Animals: Male C3H/HeN mice were purchased from Charles River Japan (Tokyo, Japan). All mice were used at the age of 7 weeks, and housed at Sankyo Laboratories (Tokyo, Japan) with free access to standard rodent chow diet.

Reagents: Lipopolysaccharide (LPS, from *Escherichia coli* O26:B6) and D-Galactosamine (GalN) were purchased from Sigma (St. Louis, MO). Enzyme-linked immunosorbent assay (ELISA) kits of murine $TNF\alpha$ were from R&D Systems (Minneapolis, MN).

TNFα Production: Naïve C3H/HeN mice (five per group) were intravenously injected with the test compound solution (10 mL/kg; dissolved in 0.1% triethylamine/saline solution), and immediately after, were intravenously injected with a mixture of LPS (0.05 mg/10 mL saline/kg) and GalN (1 g/10 mL saline/kg). Mice were injected with vehicle (0.1% triethylamine/saline solution) and saline for negative control samples, and with vehicle and LPS/GalN for positive control samples. One hour after injection, venous blood was collected under ether anesthesia with heparinized syringes fitted with 23-gauge needles from the abdominal vena, and was centrifuged at 4 °C for 3 min at $13230 \times g$ to obtain the plasma. Plasma was stored at -30 °C before measuring the TNF α levels by ELISA. The concentrations of TNF α of mouse plasma were measured using ELISA analysis according to the manufacturer's instructions.

Statistical Analysis: The percentage of inhibition of $TNF\alpha$ production was calculated by the following formula: $[1-(concentration of TNF\alpha in the test sample - concentration of <math>TNF\alpha$ in the negative control sample)/(concentration of $TNF\alpha$ in the positive control sample - concentration of $TNF\alpha$ in the negative control sample)] \times 100. The suppressive activities of test compounds are expressed as the fifty percent inhibitory concentration (IC₅₀) of the test compound, the concentration at which the test compound suppresses $TNF\alpha$ production by 50%. The IC₅₀ was calculated from the percentage of inhibition using the SAS System for Windows (version 5). The results are expressed as the mean IC₅₀ of triplicate experiments.

Production of TNFα by Mouse Peritoneal Macrophage.¹¹ C57BL/6 female mice (6-7 weeks old) were obtained from Charles River Japan, Inc., Yokohama, Japan. Peritoneal resident macrophages were collected by peritoneal lavage with ice-cold saline. After washing, cells were resuspended in the RPMI-1640 medium supplemented with 10% NBCS, 100 U/mL of penicillin and 100 µg/mL of streptomycin, and were plated in 96-well plates $(5 \times 10^4/100 \mu L/well)$. After incubation overnight at 37 °C, nonadherent cells were removed by washing three times with RPMI-1640 medium containing 10% NBCS, and adherent cells were incubated in 100 µL of the same medium, in the absence or presence of 10 ng/mL of LPS, with graded concentrations of the compounds in a humidified atmosphere of 5% CO₂ for 4.5 h at 37 °C. After incubation, the amount of TNFα produced in the culture supernatants was determined using the mouse TNF α ELISA kits. The IC₅₀ of the compounds was calculated as described above.

We thank Ms. Mariko Takai (Institute of Science and Technology, Inc.) for measuring the biological activities of the mouse peritoneal resident macrophages.

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